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Antimicrobial and Cytotoxicity Evaluation of New 3-Allyl-2-iminothiazolidin-4-ones

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Abstract

A series of novel 3-allyl-2-iminothiazolidin-4-one derivatives (**4-17**) was synthesized, through the reaction of 3-allyl-2-((3,4-dichlorophenyl)imino)thiazolidin-4-one (**3**) with different aromatic aldehydes. The chemical stability of four representative thiazolidinone derivatives (**3**, **11**, **13** and **14**) was evaluated against γ -irradiation, at a radiation dose of 25 kGy. The compounds were found to be stable with no observed degradation in liquid chromatography—mass spectrometry (LC-MS) experiments. The synthesized thiazolidinone derivatives (**3-17**) were evaluated for their antimicrobial and anticancer activities. Among the tested compounds, compounds **5**, **7**, **14** and **16** exhibited slight antibacterial activity against a multi-drug resistant *Staphylococcus aureus* (ATCC 43300 strain), at a concentration of 32 µg/mL. Compound **3** fully inhibited the growth of the fungal pathogen *Cryptococcus neoformans*, at a tested concentration of 32 µg/mL. Besides, compound **12** inhibited the biofilm formation of *S. aureus* (HG001 strain), with a percentage of 54% at a concentration of 64 µg/mL. Compared to the other tested compounds, compound **11** showed higher *in vitro* anticancer activity against melanoma and breast cancer cells, with growth inhibition values ranging from 42% to 73% at a concentration of 10 µM. Interestingly, only compounds **3**, **8** and **16** showed weak cytotoxicity to murine fibroblast L929 cells, at a tested concentration of 50 µM. The other tested compounds were not cytotoxic to fibroblasts, which suggests the relative safety of the synthesized compounds to normal mammalian cells.

Keywords: 1,3-Thiazolidin-4-one; Chemical stability; Antibacterial; Antifungal; Antibiofilm; Anticancer

1. Introduction

Infectious diseases and cancer could cause each other, as some infectious diseases interact with human cells and alter humans' essential cell signaling that could be the reason for chronic inflammation and eventually tumors formation [1]. Therefore, preventing and tackling of infectious agents have substantially affected malignant growth anticipation.

Additionally, cancer is an indirect cause of infections and the occurrence of their complications due to the deficient immunity in cancer patients, especially who are under therapeutic regimens acting on the myeloproliferative cells of the bone marrow [2].

On the other hand, infections worsen the prognosis of cancer diseases and promote metastasis [3]. Bacterial infections followed by fungal infections are the most common types of cancer-associated infections. Bacterial sepsis keeps on being the main

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cause of morbidity in pediatrics under intensive cancer treatment [4]. Furthermore, antimicrobial chemotherapy played a vital role in bacteremia prophylaxis in children with acute leukemia [5]. Therefore, there is a persistent need for more new anticancer drugs and to decrease the morbidity and mortality of microbial infections in cancer patients.

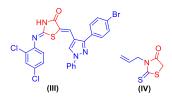
4-Thiazolidinones have been reported to be a pharmacophoric core of numerous synthetic compounds endowed with different biological actions and drug-like features [6-9]. In particular, 2substituted-1,3-thiazolidin-4-one derivatives were reported as effective antimicrobial agents [7,10,11]. 4-Thiazolidinones were also reported to have a very good ability to inhibit biofilm formation in staphylococcal bacterial strains [9,12,13]. Moreover, 2-phenylimino-4-thiazolidinones are promising compounds in the development of anticancer agents. They were reported as growth inhibitors for human lung cancer cell lines (H460 and H460/TaxR) and leukemia cell lines with lower toxicity to normal fibroblasts [14,15].

Halogenation is a pervasive approach in drug discovery and a significant number of drugs used in the clinics are halogenated [16]. Halogen atoms have a role in the interaction between proteins and their ligands through non-covalent halogen bonds [17,18]. In general, inserting halogens in molecules, such as chlorine, results in increasing the lipophilicity and consequently the biological membrane permeability without an evidence of developed toxicity [16,19]. Reported studies on the antimicrobial activity of 1,3-thiazolidinones showed also a remarkable effect of the presence of a halogenated aryl substitution at C2 [20,23].

Among the reported biologically active 1,3thiazolidin-4-ones, 2,4-dichlorophenyl the thiazolidinone derivative (I) was reported to have significant antibacterial activity against Escherichia coli, while its brominated derivative (III) showed anticancer activity against the breast cancer cell line MDA-MB-231 [20]. The alkylation of 2-phenylimino-4-thiazolidinones at N3 with small alkyl group, in particular the allyl moiety, was reported to enhance the antitumor and antimicrobial activities [24-26]. Compounds (II) and (IV), with an allyl residue at N3, were reported to exhibit significant antibacterial and antiproliferative activities, respectively [27-29]. 5-Ene-4-thiazolidinones demonstrated potent activities against various biotargets. They are potential phosphate mimics and pyrophosphate bioisosteres

[8,30]. In this study, a series of 3-allyl-5-arylidene-2-((3,4-dichlorophenyl)imino)-thiazolidin-4-one derivatives (4-17) was synthesized by introducing different arylidene moieties at C5 of the 1,3-thiazolidin-4-one ring (Figure 1).

Thiazolidinones with antibacterial activity



Thiazolidinones with anticancer activity

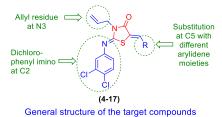


Figure 1. Reported biologically active 1,3-thaizolidin-4-ones (**I-IV**) and the design of the target compounds (**4-17**)

All the synthesized thiazolidinones 3-17 were tested for their antimicrobial (antibacterial, antibiofilm and antifungal) activity, against a panel of pathogenic bacterial and fungal strains. Five selected compounds were evaluated for their antitumor activity against 59 human-cancer cell lines. In addition, the cytotoxicity of the synthesized compounds against murine fibroblast L929 cells were evaluated. The chemical stability of four representative compounds against γ -irradiation was also studied.

2. Experimental

2.1. Chemistry

2.1.1. Materials and methods

Solvents, reagents and chemicals were bought from Alfa-Aesar and Sigma-Aldrich and used without any further purification or refining. Thin layer chromatography (TLC) plates were used to check the reactions' progress. The TLC plates were silica gel 60 F254 plated on aluminum sheets and purchased from Merck. UV light at a wave length of 254 nm was used for the visualization of the TLC plates. The used eluting solvent was a mixture of acetone/cyclohexane (3:7). Stuart apparatus was used to measure the melting points of the synthesized compounds in capillary tubes. Infrared (IR) spectroscopy for the synthesized compounds were performed by VERTEX 70 FT-IR spectrophotometer. Nuclear magnetic resonance (1H-NMR and 13C-NMR) spectra were performed in δ scale given in ppm on Varian spectrophotometer (400 MHz for ¹H-NMR and 100.6 for ¹³C-NMR) and alluded to Trimethylsilane (TMS) internal indicator. Mass spectrometry was performed by Waters ACQUITY Xevo TQD framework (Waters Corp., Milford, MA, USA), which composed of XevoTM TQD triple-quadrupole tandem mass spectrometer and ACQUITY UPLC H-Class system, with electrospray ionization (ESI) interface. The eluting solvents consisted of water containing 0.1% formic acid (solvent A) and 0.1% formic acid in acetonitrile (solvent B). Purity of the compounds was detected by HPLC (flow rate 200 µL/min).

2.1.2. Synthesis of 1-allyl-3-(3,4-dichlorophenyl)-thiourea (2)

The thiourea derivative **2** was synthesized by the reaction of 3,4-dichlorophenylisothiocynate (**1**) with allylamine, according to the reported procedure [31].

2.1.3. Synthesis of 3-allyl-2-((3,4-dichlorophenyl)-imino)thiazolidin-4-one (3)

Ethyl bromoacetate (2.5 g, 15 mmol) was added to a solution of compound $\bf 2$ (10 mmol) and anhydrous sodium acetate (0.82 g, 10 mmol) in glacial acetic acid (15 mL) or ethanol (20 mL). The reaction mixture was refluxed for 12–20 h. The solvent was evaporated and 20 mL of water was added. The formed precipitate was filtrated and crystallized from EtOH/H₂O (15:2) [32].

2.1.4. General procedure for the synthesis of compounds **4-17**

A mixture of compound **3** (0.5 mmol) and the proper aromatic aldehyde (0.6 mmol) was refluxed in 20 mL of ethanol/piperidine (30:1), for 2-12 h. The reaction was then left for cooling. The formed precipitate was filtered and crystalized from ethanol to obtain compounds **4-17**.

2.1.4.1. 3-Allyl-5-benzylidene-2-((3,4-dichlorophenyl)imino)thiazolidin-4-one (4)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with benzaldehyde. Physical form: white crystals; Yield = 90.7%; m.p. = 105-107 °C; IR, cm⁻¹: 3056 (CH aromatic), 2972, 2873 (CH aliphatic), 1707 (C=O), 1629 (C=N); 1 H-NMR (DMSO- d_6 , δ [ppm]): 4.48 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.16-5.26 (m, 2H, - $CH=CH_2$), 5.88-6.00 (m, 1H, $-CH=CH_2$), 7.02 (dd, J=2.4 and 8.4 Hz, 1H, aromatic), 7.29 (d, J = 2.4 Hz, 1H, aromatic), 7.39-7.60 (m, 5H, aromatic), 7.64 (d, J =8.4 Hz, 1H, aromatic), 7.79 (s, 1H, S-C=C<u>H</u>); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.18 (N-CH₂-), 117.77 (-CH=CH₂), 121.01 (S-C=CH), 122.04, 123.51, 127.40, 129.77 (2C), 130.37 (2C), 130.70, 131.59, 131.63, 131.89, 132.20, 133.51, 148.31, 151.77 (C=N), 165.82 (C=O); LC-MS (m/z): calculated for C₁₉H₁₄Cl₂N₂OS = 388.02, found = 389.05 [M+H]⁺; HPLC purity %: 100%.

2.1.4.2. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(4-methylbenzylidene)-thiazolidin-4-one (5)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 4-methylbenzaldehyde. Physical form: yellow needle crystals; Yield = 95.3%; m.p. = 130-132 °C; IR, cm⁻¹: 3019 (CH aromatic), 2967, 2906 (CH aliphatic), 1712 (C=O), 1623 (C=N); ¹H-NMR (DMSO-d₆, δ [ppm]): 2.31 (s, 3H, -CH₃), 4.47 (d, J = 5.2 Hz, 2H, N-CH₂), 5.16-5.24 (m, 2H, -CH=CH₂), 5.87-5.98 (m, 1H, $-CH=CH_2$), 7.02 (dd, J = 8.4 and 2.4 Hz, 1H, aromatic), 7.26-7.32 (m, 3H, aromatic), 7.44 (d, J =8.4 Hz, 2H, aromatic), 7.65 (d, J = 8.4 Hz, 1H, aromatic), 7.76 (s, 1H, S-C=CH); 13C-NMR (DMSOd₆, δ [ppm]): 21.54 (CH₃), 45.14 (N-CH₂), 117.73 (-CH=<u>C</u>H₂), 119.77 (S-<u>C</u>=CH), 122.06, 123.52, 127.35, 130.39 (2C), 130.44 (2C), 130.77, 131.67, 131.68, 131.88, 132.19, 140.95, 148.36, 151.83 (C=N), 165.89 (C=O); LC-MS (m/z): calculated for $C_{20}H_{16}Cl_2N_2OS =$ 402.04, found = 403.07 [M+H]⁺; HPLC purity %: 100%.

2.1.4.3. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(2-methoxybenzylidene)-thiazolidin-4-one (**6**)

The title compound was synthesized according to the general procedure by the reaction of compound **3** with 2-methoxybenzaldehyde. Physical form: canary yellow needle crystals; Yield = 67.8%; m.p. = 98-100 °C; IR, cm⁻¹: 3055 (CH aromatic), 2934, 2832 (CH aliphatic), 1711 (C=O), 1627 (C=N); ¹H-NMR (DMSO- d_6 , δ [ppm]): 3.86 (s, 3H, O-CH₃), 4.47 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.17-5.25 (m, 2H, -CH=C $\underline{\text{H2}}$), 5.88-5.99 (m, 1H, -C $\underline{\text{H}}$ =CH₂), 7.01 (dd, J = 8.4 and 2.4 Hz, 1H, aromatic), 7.05 (d, J = 7.2 Hz, 1H, aromatic),

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7.12 (d, J = 8.4 Hz, 1H, aromatic), 7.29 (d, J = 2.4 Hz, 1H, aromatic), 7.34 (dd, J = 7.8 and 1.4 Hz, 1H, aromatic), 7.40-7.46 (m, 1H, aromatic), 7.63 (d, J = 8.4 Hz, 1H, aromatic), 7.98 (s, 1H, S-C=C<u>H</u>); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.12 (N-CH₂-), 56.27 (O-CH₃), 112.29 (aromatic), 117.75 (-CH=<u>C</u>H₂), 120.98, 121.49, 121.99, 122.07, 123.53, 126.13, 127.33, 128.88, 131.66, 131.85, 132.16, 132.71, 148.35, 152.00 (C=N), 158.31 (2-OCH₃-<u>C</u>), 165.89 (C=O); LC-MS (m/z): calculated for C₂₀H₁₆Cl₂N₂O₂S = 418.03, found = 419.10 [M+H]⁺; HPLC purity %: 100%.

2.1.4.4. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(3-methoxybenzylidene)-thiazolidin-4-one (7)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 3-methoxybenzaldehyde. Physical form: pale yellow needle crystals; Yield = 50.3%; m.p. = 84-85 °C; IR, cm⁻¹: 3056 (CH aromatic), 2944, 2833 (CH aliph.), 1713 (C=O), 1618 (C=N); 1H-NMR (DMSO d_6 , δ [ppm]): 3.74 (s, 3H, O-CH₃), 4.47 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.17-5.24 (m, 2H, -CH=CH₂), 5.88-5.98 (m, 1H, -CH=CH₂), 6.99-7.05 (m, 2H, aromatic), 7.08 (d, J = 7.6 Hz, 1H, aromatic), 7.12 (t, J = 2.0 Hz, 1H, aromatic), 7.29 (d, J= 2.4 Hz, 1H, aromatic), 7.40 (t, J = 8.0 Hz, 1H, aromatic), 7.64 (d, J = 8.8 Hz, 1H, aromatic), 7.76 (s, 1H, S-C=CH); 13C-NMR (DMSOd₆, δ [ppm]): 45.17 (N-CH₂-), 55.71 (O-CH₃), 116.22 and 116.42 (aromatic), 117.74 (-CH=CH₂), 121.44, 121.76, 122.04, 123.51, 127.42, 130.95, 131.55, 131.59, 131.88, 132.18, 134.89, 148.22, 151.70 (C=N), 160.01 (3-OCH₃-C), 165.75 (C=O); LC-MS (m/z): calculated for $C_{20}H_{16}Cl_2N_2O_2S = 418.03$, found $= 419.08 [M+H]^+$; HPLC purity %: 100%.

2.1.4.5. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(4-methoxybenzylidene)-thiazolidin-4-one (8)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 4-methoxybenzaldehyde. Physical form: yellow needle crystals; Yield = 89.8%; m.p. = 117-118 °C; IR, cm⁻¹: 3017 (CH aromatic), 2937, 2830 (CH aliphatic), 1707 (C=O), 1626 (C=N); ¹H-NMR (DMSO-d₆, δ [ppm]): 3.78 (s, 3H, O-CH₃), 4.47 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.15-5.25 (m, 2H, -CH=CH₂), 5.88-5.99 (m, 1H, -CH=CH₂), 6.99-7.09 (m, 3H, aromatic), 7.29 (d, J = 2.4 Hz, 1H, aromatic), 7.51 (d, J = 8.8 Hz, 2H, aromatic), 7.64 (d, J = 8.8 Hz, 1H, aromatic), 7.75 (s, 1H, S-C=C<u>H</u>); 13 C-NMR (DMSO- d_6 , δ [ppm]): 45.08 (N-CH₂-), 55.87 (O-CH₃), 115.37 (2C), 117.67, 117.79, 122.11, 123.53, 126.03, 127.29, 131.60, 131.71, 131.86, 132.17, 132.44 (2C), 148.49, 152.01 (C=N), 161.28 (4-OCH₃-C), 165.98 (C=O); LC-MS

(m/z): calculated for $C_{20}H_{16}Cl_2N_2O_2S = 418.03$, found = 419.11 [M+H]⁺; HPLC purity %: 100%.

2.1.4.6. 3-Allyl-5-(3-chlorobenzylidene)-2-((3,4-dichlorophenyl)imino)-thiazolidin-4-one (9)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 3-chlorobenzaldehyde. Physical form: white flakes; Yield = 50.8%; m.p. = 113-114 °C; IR, cm⁻¹: 3074 (CH aromatic), 2972, 2930 (CH aliphatic), 1715 (C=O), 1638 (C=N); ${}^{1}\text{H-NMR}$ (DMSO- d_6 , δ [ppm]): 4.48 (d, J = 5.2 Hz, 2H, $N-CH_2$ -), 5.17-5.26 (m, 2H, - $CH=CH_2$), 5.88-5.99 (m, 1H, -CH=CH₂), 7.02 (dd, J=8.4 and 2.4 Hz, 1H, aromatic), 7.30 (d, J = 2.4 Hz, 1H, aromatic), 7.45-7.68 (m, 5H, aromatic), 7.79 (s, 1H, S-C=C<u>H</u>); 13 C-NMR (DMSO- d_6 , δ [ppm]): 45.28 (N- CH_2 -), 117.84 (- $CH=\underline{C}H_2$), 122.00 (S- $\underline{C}=CH$), 122.85, 123.50, 127.50, 127.83, 130.00, 130.30, 130.54, 131.56, 131.64, 131.91, 132.22, 134.31, 135.71, 148.14, 151.33 (C=N), 165.60 (C=O). LC-MS (*m/z*): calculated for $C_{19}H_{13}Cl_3N_2OS = 421.98$, found = 423.03 [M+H]+; HPLC purity %: 100%.

2.1.4.7. 3-Allyl-5-(4-chlorobenzylidene)-2-((3,4-dichlorophenyl)imino)-thiazolidin-4-one (10)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 4-chlorobenzaldehyde. Physical form: yellow flakes; Yield = 70.7%; m.p. = 152-153 °C; IR, cm⁻¹: 3055 (CH aromatic), 2962, 2889 (CH aliphatic), 1717 (C=O), 1640 (C=N); 1 H-NMR (DMSO- d_6 , δ [ppm]): 4.48 (d, J = 4.8 Hz, 2H, $N-CH_2$ -), 5.17-5.26 (m, 2H, - $CH=C\underline{H}_2$), 5.88-5.99 (m, 1H, $-C\underline{H}=CH_2$), 7.02 (dd, J=8.4 and 2.4 Hz, 1H, aromatic), 7.30 (d, J = 2.4 Hz, 1H, aromatic), 7.52-7.60 (m, 4H, aromatic), 7.65 (d, J =8.4 Hz, 1H, aromatic), 7.80 (s, 1H, S-C=C<u>H</u>); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.24 (N-CH₂-), 117.78 (- $CH = \underline{C}H_2$), 121.81 (S- $\underline{C} = CH$), 122.05, 123.47, 127.45, 129.85 (2C), 130.24, 131.58, 131.91, 132.04 (2C), 132.21, 132.43, 135.23, 148.28, 151.52 (C=N), 165.73 (C=O). LC-MS (m/z): calculated for C₁₉H₁₃Cl₃N₂OS = 421.98, found = 423.06 [M+H]⁺; HPLC purity %: 100%.

2.1.4.8. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(4-dimethylamino)-benzylidene)thiazolidin-4-one (11)

The title compound was synthesized according to the general procedure by the reaction of compound **3** with 4-(dimethylamino)benzaldehyde. Physical form: yellowish orange needle crystals; Yield = 88.5%; m.p. = 150-152 °C; IR, cm⁻¹: 3057 (CH aromatic), 2963, 2852 (CH aliphatic), 1707 (C=O), 1622 (C=N); ¹H-NMR (DMSO- d_6 , δ [ppm]): 2.96 (s, 6H, N(CH₃)₂), 4.45 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.15-5.23 (m, 2H, -CH=C \underline{H}_2), 5.87-5.98 (m, 1H, -C \underline{H} =CH₂), 6.77 (d, J =

9.2 Hz, 2H, aromatic), 7.02 (d, J = 8.4 and 2.4 Hz, 1H, aromatic), 7.29 (d, J = 2.4 Hz, 1H, aromatic), 7.37 (d, J = 8.8 Hz, 2H, aromatic), 7.64 (d, J = 8.4 Hz, 1H, aromatic), 7.66 (s, 1H, S-C=CH); 13 C-NMR (DMSO- d_6 , δ [ppm]): 40.66 (2C, N(CH₃)₂), 44.91 (N-CH₂-), 112.51 (2C) and 113.23 (aromatic), 117.53 (-CH=CH₂), 120.42 (S-C=CH), 122.22, 123.61, 127.08, 131.82, 131.88, 132.12, 132.44 (2C), 132.63, 148.78, 151.77, 152.45, 166.18 (C=O); LC-MS (m/z): calculated for C₂₁H₁₉Cl₂N₃OS = 431.06, found = 432.15 [M+H]⁺; HPLC purity %: 100%.

2.1.4.9. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(4-nitrobenzylidene)-thiazolidin-4-one (12)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 4-nitrobenzaldehyde. Physical form: dark yellow flakes; Yield = 85.4%; m.p. = 150-152 °C; IR, cm⁻¹: 3083 (CH aromatic), 2968, 2928 (CH aliphatic), 1712 (C=O), 1646 (C=N); ${}^{1}\text{H-NMR}$ (DMSO- d_6 , δ [ppm]): 4.50 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.18-5.229 (m, 2H, -CH=CH₂), 5.89-6.00 (m, 1H, -CH=CH₂), 7.03 (dd, J = 8.4 and 2.4 Hz, 1H, aromatic), 7.31 (d, J = 2.4 Hz, 1H, aromatic), 7.66 (d, J = 8.4 Hz, 1H, aromatic), 7.81(d, J = 8.8 Hz, 2H, aromatic), 7.90 (s, 1H, S-C=C<u>H</u>), 8.29 (d, J = 8.8 Hz, 2H, aromatic); ¹³C-NMR (DMSO d_6 , δ [ppm]): 45.39 (N-CH₂-), 117.88 (-CH=<u>C</u>H₂), 122.04 (S-<u>C</u>=CH), 123.43, 124.76 (2C), 125.54, 127.60, 128.97, 131.33 (2C), 131.48, 131.94, 132.25, 139.83, 147.76, 148.11, 151.18 (C=N), 165.51 (C=O); LC-MS (m/z): calculated for $C_{19}H_{13}Cl_2N_3O_3S =$ 433.01, found = 434.10 [M+H]^+ ; HPLC purity %: 98.47%.

2.1.4.10. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(3-(trifluoromethyl)-benzylidene)thiazolidin-4-one (13)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 3-(trifluoromethyl)benzaldehyde. Physical form: pale yellow needle crystals; Yield = 52.6%; m.p. = 150-152 °C; IR, cm⁻¹: 3061 (CH aromatic), 2980, 2935 (CH aliph.), 1711 (C=O), 1639 (C=N); ¹H-NMR (DMSO- d_6 , δ [ppm]): 4.49 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.18-5.27 (m, 2H, -CH=C<u>H</u>₂), 5.89-6.00 (m, 1H, -CH=CH₂), 7.03 (dd, J = 8.8 and 2.4 Hz, 1H, aromatic), 7.31 (d, J = 2.4 Hz, 1H, aromatic), 7.66 (d, J = 8.4 Hz, 1H, aromatic), 7.69-7.81 (m, 3H, aromatic), 7.93 (s, 1H, S-C=CH), 8.00 (s, 1H, aromatic); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.32 (N-CH₂-), 117.84 (-CH=<u>C</u>H₂), 122.03 (S-<u>C</u>=CH), 123.28, 123.48, 126.81 $(q, J= 3.9 \text{ Hz}, \text{ aromatic}), 126.92 (q, J= 269.8 \text{ Hz}, -\text{CF}_3),$ 127.52, 127.93 (q, J= 3.9 Hz), 129.95, 130.37 (q, J= 32.2 Hz, C-CF₃), 131.00, 131.55, 131.91, 132.22, 132.60, 134.69, 148.12, 151.26 (C=N), 165.58 (C=O); LC-MS (m/z): calculated for $C_{20}H_{13}Cl_2F_3N_2OS =$

456.01, found = 457.12 [M+H]⁺; HPLC purity %: 100%.

2.1.4.11. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(2,4-dimethoxy-benzylidene)thiazolidin-4-one (14)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 2,4-dimethoxybenzaldehyde. Physical form: canary yellow flakes; Yield = 69.8%; m.p. = 123-124 °C; IR, cm⁻¹: 3017 (CH aromatic), 2933, 2857 (CH aliphatic), 1707 (C=O), 1628 (C=N); ¹H-NMR (DMSO- d_6 , δ [ppm]): 3.79 (s, 3H, 4-OCH₃), 3.86 (s, 3H, 2-OCH₃), 4.45 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.16-5.23 (m, 2H, $-CH=C\underline{H}_2$), 5.5.87-5.98 (m, 1H, - $CH = CH_2$, 6.62-6.68 (m, 2H, aromatic), 7.01 (dd, J =8.6, 2.6 Hz, 1H, aromatic), 7.25-7.30 (m, 2H, aromatic), 7.63 (d, J = 8.4 Hz, 1H, aromatic), 7.92 (s, 1H, S-C=CH); ¹³C-NMR (DMSO-*d*₆, δ [ppm]): 45.01 (N-CH₂-), 56.04 (4-OCH₃), 56.42 (2-OCH₃), 99.05, 107.01 and 114.83 (aromatic), 117.47, 117.65, 122.14, 123.55, 126.08, 127.21, 130.33, 131.74, 131.83, 132.13, 148.53, 152.24 (S-C=N), 160.10 (2-OCH₃-C), 163.33 (4-OCH₃-C), 166.09 (C=O); LC-MS (*m/z*): calculated for $C_{21}H_{18}Cl_2N_2O_3S = 448.04$, found = 449.16 [M+H]+; HPLC purity %: 100%.

2.1.4.12. 3-Allyl-5-(3,4-dichlorobenzylidene)-2-((3,4-dichlorophenyl)imino)-thiazolidin-4-one (15)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 3,4-dichlorobenzaldehyde. Physical form: pale buff flakes; Yield = 55.7%; m.p. = 143-144 °C; IR, cm⁻ 1: 3072 (CH aromatic), 2980, 2935 (CH aliphatic), 1714 (C=O), 1635 (C=N); 1 H-NMR (DMSO- d_6 , δ [ppm]): 4.48 (d, J = 5.2 Hz, 2H, N-CH₂-), 5.17-5.27 (m, 2H, $-CH=C\underline{H}_2$), 5.88-5.99 (m, 1H, $-C\underline{H}=CH_2$), 7.02 (dd, J = 8.8 and 2.4 Hz, 1H, aromatic), 7.30 (d, J= 2.4 Hz, 1H, aromatic), 7.48 (dd, J = 8.4 and 2.0 Hz, 1H, aromatic), 7.66 (d, J = 8.4 Hz, 1H, aromatic), 7.73 (d, J = 8.4 Hz, 1H, aromatic), 7.79 (s, 1H, S-C=C<u>H</u>), 7.90 (d, J = 2.0 Hz, 1H, aromatic); ¹³C-NMR (DMSO d_6 , δ [ppm]): 45.33 (N-CH₂-), 117.85 (-CH=<u>C</u>H₂), 122.01 (S-C=<u>C</u>H), 123.39, 123.48, 127.54, 128.97, 128.99, 131.53, 131.93 (2C), 132.24, 132.40, 132.84, 132.94, 134.29, 148.13, 151.17 (C=N), 165.54 (C=O); LC-MS (m/z): calculated for C₁₉H₁₂Cl₄N₂OS = 455.94, found = $457.05 [M+H]^+$; HPLC purity %: 98.41%.

2.1.4.13. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(3,4,5-trimethoxy-benzylidene)thiazolidin-4-one (**16**)

The title compound was synthesized according to the general procedure by the reaction of compound **3** with 3,4,5-trimethoxybenzaldehyde. Physical form: canary yellow needle crystal; Yield = 43.7%; m.p. = 102-104 °C; IR, cm⁻¹: 3063 (CH aromatic), 2930, 2833 (CH aliphatic), 1707 (C=O), 1629 (C=N); ¹H-NMR

(DMSO- d_6 , δ [ppm]): 3.69 (s, 3H, 4-OCH₃), 3.75 (s, 6H, 3,5-dimethoxy), 4.48 (d, J= 5.2 Hz, 2H, N-CH₂-), 5.19-5.25 (m, 2H, -CH=C<u>H</u>₂), 5.88-5.99 (m, 1H, -C<u>H</u>=CH₂), 6.86 (s, 2H, aromatic), 7.05 (dd, J = 8.4 and 2.4 Hz, 1H, aromatic), 7.33 (d, J = 2.4 Hz, 1H, aromatic), 7.63 (d, J = 8.4 Hz, 1H, aromatic), 7.76 (s, 1H, S-C=C<u>H</u>); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.17 (N-CH₂-), 56.55 (2C, 3,5-dimethoxy), 60.66 (4-OCH₃), 108.05 (2C, aromatic), 117.70 (-CH=<u>C</u>H₂), 120.13 (S-<u>C</u>=CH), 122.12, 123.63, 127.34, 129.19, 131.66, 131.79, 131.95, 132.13, 139.89, 147.96, 151.50, 153.62 (2C, (3,5-dimethoxy)<u>C</u>₂), 165.69 (C=O); LC-MS (m/z): calculated for C₂₂H₂₀Cl₂N₂O₄S = 478.05, found = 479.13 [M+H]⁺; HPLC purity %: 99.02%.

2.1.4.14. 3-Allyl-2-((3,4-dichlorophenyl)imino)-5-(thiophen-2-ylmethylene)-thiazolidin-4-one (17)

The title compound was synthesized according to the general procedure by the reaction of compound 3 with 2-thiophenecarboxaldehyde. Physical form: canary yellow flakes; Yield = 78.4%; m.p. = 117-118 °C; IR, cm⁻¹: 3076 (CH aromatic), 2937, 2889 (CH aliphatic), 1710 (C=O), 1631 (C=N); ¹H-NMR (DMSO- d_6 , δ [ppm]): 4.47 (d, J= 5.2, 2H, N-CH₂-), 5.16-5.25 (m, 2H, -CH=C \underline{H}_2), 5.87-5.99 (m, 1H, - $CH = CH_2$, 7.04 (dd, J = 8.8 and 2.4 Hz, 1H, aromatic), 7.24 (dd, J = 5.0 and 3.8 Hz, 1H, aromatic), 7.32 (d, J= 2.4 Hz, 1H, aromatic), 7.64 (d, J = 3.6 Hz, 1H, aromatic), 7.67 (d, J = 8.8 Hz, 1H, aromatic), 7.91 (d, J = 4.8 Hz, 1H, aromatic), 8.06 (s, 1H, S-C=CH); ¹³C-NMR (DMSO- d_6 , δ [ppm]): 45.29 (N-CH₂-), 117.72 (- $CH = \underline{C}H_2$), 118.44 (S- \underline{C} =CH), 122.10, 123.54, 124.93, 127.44, 129.35, 131.65, 131.90, 132.19, 133.20, 134.71, 137.64, 148.38, 151.37 (C=N), 165.59 (C=O); LC-MS (m/z): calculated for $C_{17}H_{12}Cl_2N_2OS_2 =$ 393.98, found = 395.03 [M+H]+; HPLC purity %:

2.2. Irradiation of the synthesized compounds

The tested compounds, in solid form, were collected in polypropylene vials and wrapped with aluminum sheet. The vials were subjected to γ -radiation dose of 25 kGy. γ -Irradiation was performed by using a ⁶⁰Co source, with a dose rate of 1.119 kGy/h, utilizing Indian-Gamma Cell (Ge 4000 A).

2.3. Biological evaluation

2.3.1. Screening against bacterial and fungal strains

The antimicrobial activity was tested against 5 bacterial strains (*Staphylococcus aureus* (MRSA) ATCC 43300, *E. coli* ATTCC 25922, *Klebsiella pneumonia* ATCC 700603, *Pseudomonas aeruginosa*

ATCC 27853 and Acinetobacter baumannii ATCC 19606) and 2 fungal strains (Candida albicans ATCC 90028 and Cryptococcus neoformans ATCC 208821). The antimicrobial screening was performed by the Community for Antimicrobial Drug Discovery (CO-ADD), funded by the Wellcome Trust (United Kingdom) and University of Queensland (Australia).

2.3.2. Inhibition of biofilm formation

The synthesized compounds were evaluated for their biofilm inhibition activity in a microtiter format utilizing a reported procedure [33]. Overnight culture of S. aureus (HG001) was diluted 1:100 in 0.5X Tryptic soy broth appended with 1% glucose (culture medium). Compounds' solutions in DMSO and bacterial suspensions were added in the wells of the 96-well plates. The final concentration of DMSO in each well would be \leq 3%. The covered plates were incubated at 37 °C for 24 hours in the incubator. The optical density of the overall bacterial growth was measured spectrophotometrically at 630 nm for each well. The plates wells were washed by using a washer device (Biotek ELx50) after the removal of the bacterial suspension. 50 µL of watery 0.06% (w/v) crystal violet solution to each well to stain the attached biofilm. The solution was removed, and the wells were washed three times with distilled water and dried, followed by the addition of 200 μL of acetic acid (30%) in each well to elute crystal violet. Direct quantification of the biofilm by using microplate reader (Biotek ELx800) at 630 nm. The data are presented as per cent inhibition of S. aureus (HG001) biofilm for the synthesized compounds prorated to the negative control (DMSO).

2.3.3. Anticancer evaluation

Five compounds (4, 11, 13, 14 and 17) were selected by the National Cancer Institute (NCI), Bethesda, MD, USA, for evaluating their anticancer activity. The selected compounds were subjected to a primary *in vitro* one-dose (10 μ M) assay against 59 human tumor cell lines. The cell lines used in the screening included leukemia, non-small cell lung cancer, colon cancer, CNS cancer, melanoma, ovarian cancer, renal cancer, prostate cancer and breast cancer cells.

2.3.4. Cytotoxicity evaluation against murine fibroblast (L929) cells

The compounds were screened for their cytotoxicity using the alamarBlue reagent [ThermoFischer Scientific], a resazurin based solution to measure cell viability. L929 cells were cultured in Dulbecco's Modified Eagle Medium (DMEM)

supplemented with 10% fetal bovine serum (FBS) and 1.56*104 cells/mL was seeded in 96-well cell culture plates with a total volume of 60 µL. The plates with cells were incubated for 24 h at 37 °C and 10 % CO₂. Subsequently, 0.6 µL of compound solutions were added per well (for final a concentration of 100 µM). The plate was placed on a plate shaker for 15 seconds at 1000 rpm to ensure the optimal mixing. After 72 h of incubation at 37 °C and 10 % CO₂, 5 µL of alamarBlue reagent was added to each well. The fluorescence intensity of each well was determined after 2.5 h of incubation at 37 °C and 10 % CO₂, using an extinction wavelength of (\lambda max, abs= 540 nm) and an emission wavelength of 600 nm (λmax, em= 600 nm). For data analysis, the mean value and standard deviation were determined from three replicates. DMSO and water were used as vehicle controls; cells without compounds and pure medium without cells and compounds as negative controls. The final concentration of DMSO in each well did not exceed 1%. Compounds 3, 5, 8, 11, 14, 16 and 17 were further tested at concentrations of 50, 30, 12.5, 5, 1.3 and 0.4 μΜ.

3. Results and discussion

3.1. Chemistry

The synthetic route of compounds 2-17 is illustrated in Scheme 1. The reported thiourea derivative 2 was obtained by nucleophilic addition reaction of 3,4-dichlorophenyl-isothiocyanate (1) with allylamine, according to the reported method [24, 31, 32]. The next step was S_N2 nucleophilic substitution rection of the thiourea derivative (2) with ethyl bromoacetate, catalyzed by the presence of sodium acetate anhydrous, followed by intramolecular cyclization reaction to afford the previously reported thiazolidinone derivative (3) [32, 34]. The preparation of thiazolidinone derivative 3 was performed in glacial acetic acid, or in absolute ethanol where the reaction yield was higher. The target 5-ene-4-thiazolidinones (4-17) were obtained as pure crystals through a Knoevenagel condensation reaction of the active methylene (C5) of compound 3 with the carbonyl group of the corresponding aldehyde in a mixture of ethanol and piperidine (30:1) [35].

IR spectroscopy, ¹H-NMR, ¹³C-NMR and LC-MS were used to confirm the structures of the target compounds. FT-IR spectra of compounds (**4-17**) revealed a shift in the value of the carbonyl bands from 1760 cm⁻¹ to a range of 1707-1717 cm⁻¹. This bathochromic shift is due to the conjugation of the carbonyl group with the arylidene moiety [36]. ¹H-NMR spectra of compounds **4-17** showed signals for

the one olefinic proton (S-C=CH) at δ ranging from 7.6 to 8 ppm, which confirms the occurrence of the reaction between compound 3 and the corresponding aldehydes. Therefore, the exocyclic C=C bond of the synthesized thiazolidinone derivatives (4-17) was confirmed to be in Z-configuration by those singlet signals of olefinic protons of all derivatives, which resonate at higher chemical shifts due to the deshielding effect of the adjacent carbonyl group. Otherwise, the olefinic proton would resonate towards the right at δ value less than 6.64 ppm if it were in *E*-configuration [37,38].

NCS
$$CI \longrightarrow H_2N \longrightarrow CI \longrightarrow CI$$

$$1 \longrightarrow CI \longrightarrow CI$$

$$2 \longrightarrow CI \longrightarrow Br \longrightarrow O$$

$$N \longrightarrow S$$

$$R \longrightarrow H$$

$$CI \longrightarrow CI$$

Compound	R	Compound	R	
4		11	}-N	
5	*	12	₩ NO ₂	
6	-0	13	CF ₃	
7	₽————————————————————————————————————	14	-0-	
8	₩	15	CI	
9	CI	16	0-	
10	€—CI	17	S	

Scheme 1. Synthesis of compounds 2-17. Reagents and conditions: (i) Toluene, room temperature, 1 h or ethanol, reflux, 6 h (ii) Sodium acetate anhydrous, ethyl bromoacetate, ethanol or glacial acetic acid, reflux, 20 h (iii) Ethanol/piperidine (30:1), reflux 2-12 h.

The purity and the molecular masses of the synthesized compounds were tested by LC-MS analyses. With the purpose of evaluating the chemical stability of the thiazolidinone derivatives (3-17), four compounds (3, 11, 13 and 14) were selected and exposed to y-irradiation at single dose of 25 kGy [39-42]. The physical and the chemical properties of these tested compounds, in their solid state, were studied before and after the irradiation process. The results revealed no changes in the physical and the chemical properties, including the color, form and solubility. Thin layer chromatography (TLC) experiments showed no change in the R_f values of the tested compounds and no additional spots were observed. In addition, LC-MS experiments were performed for the four tested compounds (3, 11, 13 and 14), before and after irradiation. The results of the LC-MS experiments revealed no change in the mass nor in the purity of any of the tested compounds, which indicates the chemical stability of the synthesized compounds.

3.2. Biological evaluation

All the target compounds (3-17) were tested for their *in vitro* antibacterial activity against five pathogenic bacterial strains (Table 1). The used bacterial strains included the Gram-positive multidrug resistant *S. aureus* (MRSA) ATCC 43300, and the Gram-negative bacteria *E. coli* ATTCC 25922, *K.*

pneumoniae ATCC 700603, *P. aeruginosa* ATCC 27853 and *A. baumannii* ATCC 19606. Compounds **5**, **7**, **14** and **16** exhibited a slight antibacterial activity against MRSA, with a growth inhibition percentage of 21%, 24%, 26% and 23% at a concentration of 32 μg/mL, respectively. It was clearly observed that the presence of a methyl group (compound **5**) and methoxy groups (in compounds **7**, **14** and **16**), in the meta and para positions, enhanced the antibacterial activity of the tested compounds against MRSA. The other compounds showed lower activity against the used Gram-positive and Gram-negative bacteria (Table 1).

The 1,3-thiazolidine-4-one synthesized derivatives (3-17) were also evaluated for their antifungal activity against C. albicans ATCC 90028 and C. neoformans var. grubii H99; ATCC 208821 (Table 1). Compound 3 exhibited a potent in vitro antifungal activity against C. neoformans, with a full growth inhibition (100% inhibition), at a concentration of 32 µg/mL. The other tested compounds displayed weak or no antifungal activity against the tested fungal strains. The results of the antifungal screening confirmed that the unsubstituted C5 of the thiazolidinone ring is essential for the antifungal activity against C. neoformans. Insertion of substitutions on C5 diminished the antifungal activity.

Table 1. Antibacterial, antifungal and antibiofilm activities of the synthesized compounds

C	Antibacterial activity Growth inhibition percent (GI%) of bacterial strains, at a concentration of 32 µg/mL			Antifungal activity Growth inhibition percent (Gl%) of fungal strains, at a concentration of 32 µg/mL		Anti-biofilm activity Inhibition percent of biofilm formation of S. aureus HG001			
Compound	MRSA ATCC 43300	E. coli ATTCC 25922	K. pneumniae ATCC 700603	P. aeruginosa ATCC 27853	A. baumannii ATCC 19606	C. albicans ATCC 90028	C. neoformans Var. grubii H99; ATCC 208821	32 μg/mL	64 μg/mL
3	<10	<10	<10	<10	<10	10	100	<10	<10
4	15	14	11	11	<10	12	19	<10	<10
5	21	17	11	13	<10	<10	18	<10	<10
6	16	12	<10	11	<10	<10	18	<10	<10
7	24	18	<10	13	<10	<10	17	<10	22
8	19	15	16	<10	<10	<10	13	<10	<10
9	<10	<10	<10	<10	<10	<10	<10	<10	19
11	<10	12	<10	10	<10	<10	<10	<10	13
12	18	16	11	<10	13	<10	<10	35	54
13	10	<10	<10	<10	<10	10	<10	<10	26
14	26	16	13	15	22	<10	<10	<10	<10
15	17	16	20	<10	<10	<10	<10	<10	<10
16	23	20	19	12	16	<10	<10	<10	<10

S. aureus belongs to "ESKAPE" pathogens; (IC₅₀ values ranging from 80.32 to $>250 \mu M$), which

pathogenic bacteria with the highest impact in bacterial resistance [43]. In many cases, persistent infections caused by *S. aureus* are due to its high capability to resist antibiotics via biofilm formation [44]. Therefore, new molecules capable of defeating bacterial biofilm are needed to overcome resistant infections. The ability of the synthesized compounds to inhibit the biofilm formation of *S. aureus* HG001 was also evaluated. Among the tested compounds, the 3-allyl-2-iminothiazolidin-4-one derivative 12 inhibited biofilm formation with percentages of 35% and 54% at tested concentrations of 32 and 64 µg/mL, respectively (Table 1).

Five 5-(arylidene)thiazolidin-4-one derivatives (compounds 4, 11, 13, 14 and 17) were selected by the NCI (National Cancer Institute - Developmental Therapeutic Program), to be evaluated for their in vitro anticancer activity. The five selected compounds were tested against a panel of 59 human-cancer cell lines, including leukemia, non-small cell lung cancer, ovarian cancer, CNS cancer, renal cancer, melanoma, prostate cancer, colon cancer and breast cancer cells (Table S1 in the supplementary file). Out of the used 59 cell lines, the five tested compounds exhibited anticancer activity against 18 cell lines, as presented in Table 2. The 5-(4-dimethyl-aminobenzylidene)thiazolidin-4-one derivative 11 was the most active compound. Compound 11, bearing a dimethylamino moiety, inhibited the growth of MDA-MB-435, UACC-62 and MCF7 cells with growth inhibition values of 73%, 45% and 42%, respectively, at concentration of 10 µM. The other tested compounds showed weaker anticancer activity, with growth inhibition values ranging from < 20% up to 47%.

Cytotoxicity of compounds 3-17 was tested against murine fibroblast L929 cells. The test was performed at a concentration of 100 μ M with an incubation period of 3 days. Cell viability was measured using alamarBlue reagent method. Only compounds 3, 5, 8, 11, 14, 16 and 17 showed slight cytotoxicity at the tested concentration (100 μ M), while the other tested compounds did not show cytotoxicity to murine fibroblast L929 cells. Compounds 3, 5, 8, 11, 14, 16 and 17 were further tested to calculate their IC₅₀ values. No significant cytotoxicity was observed for the tested compounds

Table 2. Growth inhibition percent (GI%) of human-cancer cell lines, at a concentration of $10 \mu M$

suggests their relative safety on fibroblasts (Table 3).

Cell line			Compound				
		4	11	13	14	17	
Leukaemia	SR	<20	29	<20	<20	<20	
Non-Small	HOP-92	<20	25	20	<20	<20	
Cell Lung Cancer	NCI-H522	<20	32	23	<20	24	
	HCT-116	<20	<20	32	<20	<20	
	HCT-15	<20	26	28	<20	<20	
Colon Cancer	HT29	<20	<20	33	<20	20	
	KM12	<20	20	<20	<20	<20	
	SW-620	<20	22	<20	<20	<20	
CNS Cancer	SNB-75	<20	<20	<20	25	<20	
	MALME-3M	<20	24	<20	<20	<20	
Melanoma	MDA-MB-435	<20	73	<20	<20	<20	
	UACC-62	21	45	31	20	25	
Renal Cancer	A498	<20	35	<20	<20	30	
	UO-31	24	31	<20	22	<20	
Prostate Cancer	PC-3	<20	23	<20	<20	<20	
	MCF7	<20	42	47	<20	20	
Breast Cancer	BT-549	<20	22	<20	<20	<20	
	T-47D	24	<20	<20	<20	<20	

Table 3. Cytotoxicity against murine fibroblast (L929) cells

Compound	Growth inhibition % at 50 μM	Calculated IC ₅₀ in µM
3	34.25	94.3
5	10.62	214.14
8	25.41	80.32
11	8.89	>250
14	8.29	146.6
16	26.08	95.61
17	14.4	225.9

4. Conclusion

We report the synthesis of new halogenated 3-allyl-2-iminothiazolidin-4-ones (4-17). The synthesized compounds were subjected to

antibacterial, antifungal, anti-biofilm formation and anticancer screening. Four compounds (5, 7, 14 and 16) exhibited slight antibacterial activity. The common feature of the four compounds 5, 7, 14 and 16 was the presence of methyl or methoxy groups in the meta or the para positions of the benzylidene group. Compound 3, with no substitution at C5 of the 1,3thiazolidine-4-one, caused a complete inhibition of the growth of the fungal pathogen C. neoformans. The antifungal activity of compound 3 proves that unsubstituted C5 is important for the antifungal activity. It was also clearly observed that compound 11, with a dimethylamino group at the para position, has a remarkable in vitro anticancer activity. Melanoma cells were the most sensitive cancer cells to compound 11, followed by breast cancer cells. The other tested compounds showed lower anticancer activity. Moreover, all the synthesized thiazolidinone derivatives did not show significant cytotoxicity to murine fibroblast cells. Taken together, the results of this study showed a clear relationship between the structures and the biological activity of the tested compounds. Consequently, these results could be a guide to synthesize more 1,3-thaizolidin-4-one derivatives, in order to develop new more active compounds.

5. Acknowledgement

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6. Conflict of interest

The authors declare no conflict of interest.

7. References

- 1. van Elsland D. and Neefjes J., Bacterial infections and cancer., **19**(11), e46632 (2018).
- Zorina T. and Styche A., Infectious Diseases in Cancer Patients: An Overview, in Infection and Cancer: Bi-Directorial Interactions, M.R. Shurin, Y. Thanavala, and N. Ismail, Editors. Springer International Publishing: Cham. p. 295-311 (2015).
- Attiê R., Chinen L.T.D., Yoshioka E.M., Silva M.C.F. and de Lima V.C.C., Acute bacterial infection negatively impacts cancer specific survival of colorectal cancer patients. World Journal of Gastroenterology, 20(38), 13930-13935 (2014).
- Alexander S., Nieder M., Zerr D.M., Fisher B.T., Dvorak C.C. and Sung L., Prevention of bacterial infection in pediatric oncology: What do we know, what can we learn? *Pediatric Blood & Cancer*, 59(1), 16-20 (2012).
- Alexander S., Fisher B.T., Gaur A.H., Dvorak C.C., Villa Luna D., Dang H., Chen L., Green M., Nieder M.L., Fisher B., Bailey L.C., Wiernikowski J., Sung L. and f.t.C.s.O. Group, Effect of Levofloxacin Prophylaxis on Bacteremia in Children With Acute Leukemia or Undergoing Hematopoietic Stem Cell Transplantation: A Randomized Clinical Trial. *JAMA*, 320(10), 995-1004 (2018).
- Lesyk R., Drug design: 4-thiazolidinones applications. Part 1. Synthetic routes to the drug-like molecules. *Journal of Medical Science*, 89(1), e406 (2020).
- 7. Kaur Manjal S., Kaur R., Bhatia R., Kumar K., Singh V., Shankar R., Kaur R. and Rawal R.K., Synthetic and medicinal perspective of thiazolidinones: A review. *Bioorganic Chemistry*, **75**, 406-423 (2017).
- 8. Kaminskyy D., Kryshchyshyn A. and Lesyk R., 5-Ene-4-thiazolidinones – An efficient tool in medicinal chemistry. *European Journal of Medicinal Chemistry*, **140**, 542-594 (2017).
- 9. El-Hossary E.M., Nissan Y.M., Edkins K. and Bruhn H., Synthesis and antibacterial activity of novel 2-(arylimino)thiazolidin-4-one and 2-(benzylidenehydrazono)-3-arylthiazolidin-4-one derivatives. *Journal of Applied Pharmaceutical Science*, **6**(5), 7-17 (2016).
- Verma A. and Saraf S.K., 4-Thiazolidinone A biologically active scaffold. European Journal of Medicinal Chemistry, 43(5), 897-905 (2008).
- 11. Tripathi A.C., Gupta S.J., Fatima G.N., Sonar P.K., Verma A. and Saraf S.K., 4-Thiazolidinones: The advances continue.... *European Journal of Medicinal Chemistry*, **72**, 52-77 (2014).
- 12. Rane R.A., Sahu N.U. and Shah C.P., Synthesis and antibiofilm activity of marine natural product-based 4-

- thiazolidinones derivatives. *Bioorganic & Medicinal Chemistry Letters*, **22**(23), 7131-7134 (2012).
- 13. Angapelly S., Sri Ramya P.V., SunithaRani R., Kumar C.G., Kamal A. and Arifuddin M., Ultrasound assisted, VOSO4 catalyzed synthesis of 4-thiazolidinones: Antimicrobial evaluation of indazole-4-thiazolidinone derivatives. *Tetrahedron Letters*, **58**(49), 4632-4637 (2017).
- Zhou H., Wu S., Zhai S., Liu A., Sun Y., Li R., Zhang Y., Ekins S., Swaan P.W., Fang B., Zhang B. and Yan B., Design, Synthesis, Cytoselective Toxicity, Structure–Activity Relationships, and Pharmacophore of Thiazolidinone Derivatives Targeting Drug-Resistant Lung Cancer Cells. *Journal of Medicinal Chemistry*, 51(5), 1242-1251 (2008).
- Senkiv J., Finiuk N., Kaminskyy D., Havrylyuk D., Wojtyra M., Kril I., Gzella A., Stoika R. and Lesyk R., 5-Ene-4-thiazolidinones induce apoptosis in mammalian leukemia cells. *European Journal of Medicinal Chemistry*, 117, 33-46 (2016).
- Marcelo Zaldini H., Suellen Melo T.C., Diogo Rodrigo M.M., Walter Filgueira de Azevedo J. and Ana Cristina Lima L., Halogen Atoms in the Modern Medicinal Chemistry: Hints for the Drug Design. Current Drug Targets, 11(3), 303-314 (2010).
- Gilday L.C., Robinson S.W., Barendt T.A., Langton M.J., Mullaney B.R. and Beer P.D., Halogen Bonding in Supramolecular Chemistry. *Chemical Reviews*, 115(15), 7118-7195 (2015).
- Cavallo G., Metrangolo P., Milani R., Pilati T., Priimagi A., Resnati G. and Terraneo G., The Halogen Bond. *Chemical Reviews*, 116(4), 2478-2601 (2016).
- Gillis E.P., Eastman K.J., Hill M.D., Donnelly D.J. and Meanwell N.A., Applications of Fluorine in Medicinal Chemistry. *Journal of Medicinal Chemistry*, 58(21), 8315-8359 (2015).
- Bhat M., Poojary B., Kalal B.S., Swamy P.M.G., Kabilan S., Kumar V., Shruthi N., Anand S.A.A. and Pai V.R., Synthesis and evaluation of thiazolidinone– pyrazole conjugates as anticancer and antimicrobial agents. *Future Medicinal Chemistry*, 10(9), 1017-1036 (2018).
- Brahmbhatt H., Bhatt A.K., Das A.K., Paul P. and Sharma S., 2-(3,4-Dichlorophenylimino)-5-((3-(p-substitutedphenyl)-1-phenyl-1*H*-pyrazol-4-yl)methylene)thiazolidin-4-one as an Antibacterial, Antifungal and Antimycobacterial Agent. *Journal of Heterocyclic Chemistry*, 54(5), 2838-2843 (2017).
- Chawla P., Singh R. and Saraf S.K., Effect of chloro and fluoro groups on the antimicrobial activity of 2,5disubstituted 4-thiazolidinones: a comparative study. *Medicinal Chemistry Research*, 21(10), 3263-3271 (2012).
- 23. Bondock S., Khalifa W. and Fadda A.A., Synthesis and antimicrobial evaluation of some new thiazole, thiazolidinone and thiazoline derivatives starting from 1-chloro-3,4-dihydronaphthalene-2-carboxaldehyde. *European Journal of Medicinal Chemistry*, **42**(7), 948-954 (2007).
- Ottanà R., Carotti S., Maccari R., Landini I., Chiricosta G., Caciagli B., Vigorita M.G. and Mini E.,

- *In vitro* antiproliferative activity against human colon cancer cell lines of representative 4-thiazolidinones. Part I. *Bioorganic & Medicinal Chemistry Letters*, **15**(17), 3930-3933 (2005).
- Kawakami M., Koya K., Ukai T., Tatsuta N., Ikegawa A., Ogawa K., Shishido T. and Chen L.B., Structure–Activity of Novel Rhodacyanine Dyes as Antitumor Agents. *Journal of Medicinal Chemistry*, 41(1), 130-142 (1998).
- Azizmohammadi M., Khoobi M., Ramazani A., Emami S., Zarrin A., Firuzi O., Miri R. and Shafiee A., 2H-chromene derivatives bearing thiazolidine-2,4dione, rhodanine or hydantoin moieties as potential anticancer agents. European Journal of Medicinal Chemistry, 59, 15-22 (2013).
- 27. Karalı N., Terzioğlu N., and Gürsoy A., Synthesis and Primary Cytotoxicity Evaluation of New 5-Bromo-3-substituted-hydrazono-1*H*-2-indolinones. *Archiv der Pharmazie*, **335**(8), 374-380 (2002).
- 28. Nasr T., Bondock S. and Eid S., Design, synthesis, antimicrobial evaluation and molecular docking studies of some new 2,3-dihydrothiazoles and 4-thiazolidinones containing sulfisoxazole. *Journal of Enzyme Inhibition and Medicinal Chemistry*, **31**(2), 236-246 (2016).
- 29. Ali Muhammad S., Ravi S. and Thangamani A., Synthesis and evaluation of some novel N-substituted rhodanines for their anticancer activity. *Medicinal Chemistry Research*, **25**(5), 994-1004 (2016).
- Holota S., Kryshchyshyn A., Derkach H., Trufin Y., Demchuk I., Gzella A., Grellier P. and Lesyk R., Synthesis of 5-enamine-4-thiazolidinone derivatives with trypanocidal and anticancer activity. *Bioorganic Chemistry*, 86, 126-136 (2019).
- 31. Ramadas K., Suresh G., Janarthanan N. and Masilamani S., Antifungal activity of 1,3-disubstituted symmetrical and unsymmetrical thioureas. *Pesticide Science*, **52**(2), 145-151 (1998).
- 32. Hammad S.G., El-Gazzar M.G., Abutaleb N.S., Li D., Ramming I., Shekhar A., Abdel-Halim M., Elrazaz E.Z., Seleem M.N., Bilitewski U., Abouzid K.A.M. and El-Hossary E.M., Synthesis and antimicrobial evaluation of new halogenated 1,3-Thiazolidin-4-ones. *Bioorganic Chemistry*, **95**, 103517 (2020).
- 33. Kwasny S.M. and Opperman T.J., Static Biofilm Cultures of Gram-Positive Pathogens Grown in a Microtiter Format Used for Anti-Biofilm Drug Discovery. *Current Protocols in Pharmacology*, **50**(1), 13A.8.1-13A.8.23 (2010).
- 34. Yella R., Ghosh H. and Patel B.K., It is "2-imino-4-thiazolidinones" and not thiohydantoins as the reaction product of 1,3-disubstituted thioureas and chloroacetylchloride. *Green Chemistry*, **10**(12), 1307-1312 (2008).
- Poyraz Ö., Jeankumar V.U., Saxena S., Schnell R., Haraldsson M., Yogeeswari P., Sriram D. and Schneider G., Structure-Guided Design of Novel Thiazolidine Inhibitors of O-Acetyl Serine Sulfhydrylase from Mycobacterium tuberculosis. Journal of Medicinal Chemistry, 56(16), 6457-6466 (2013).

- Seshadri T.R., Rao N.V.S. and Subrahmanyam B., Effect of conjugation and complex formation on the Raman and I.R. frequencies of the carbonyl group. Proceedings of the Indian Academy of Sciences -Section A, 68(6), 314-323 (1968).
- 37. Mushtaque M., Avecilla F. and Azam A., Synthesis, characterization and structure optimization of a series of thiazolidinone derivatives as *Entamoeba histolytica* inhibitors. *European Journal of Medicinal Chemistry*, **55**, 439-448 (2012).
- Ottanà R., Maccari R., Barreca M.L., Bruno G., Rotondo A., Rossi A., Chiricosta G., Di Paola R., Sautebin L., Cuzzocrea S. and Vigorita M.G., 5-Arylidene-2-imino-4-thiazolidinones: Design and synthesis of novel anti-inflammatory agents. *Bioorganic & Medicinal Chemistry*, 13(13), 4243-4252 (2005).
- Jacobs G.P., A Review of the Effects of Gamma Radiation on Pharmaceutical Materials. *Journal of Biomaterials Applications*, 10(1), 59-96 (1995).
- 40. Crucq A.-S., Deridder V. and Tilquin B., Radiostability of pharmaceuticals under different irradiation conditions. *Radiation Physics and Chemistry*, **72**(2), 355-361 (2005).
- 41. Singh B.K., Parwate D.V., Das Sarma I.B. and Shukla S.K., Study on gamma and electron beam sterilization of third generation cephalosporins cefdinir and cefixime in solid state. *Radiation Physics and Chemistry*, **79**(10), 1079-1087 (2010).
- Zalewski P., Skibiński R., Szymanowska-Powałowska D., Piotrowska H., Bednarski W. and Cielecka-Piontek J., Radiolytic studies of cefozopran hydrochloride in the solid state. *Electronic Journal of Biotechnology*, 25, 28-32 (2017).
- 43. Pendleton J.N., Gorman S.P. and Gilmore B.F., Clinical relevance of the ESKAPE pathogens. *Expert Review of Anti-infective Therapy*, **11**(3), 297-308 (2013)
- 44. Neopane P., Nepal H.P., Shrestha R., Uehara O. and Abiko Y., *In vitro* biofilm formation by Staphylococcus aureus isolated from wounds of hospital-admitted patients and their association with antimicrobial resistance. *International Journal of General Medicine*, 11, 25-32 (2018).