



Qualitative And Quantitative Properties Of Essential Oil Of *Mentha Pulegium* L. And *Mentha Suaveolens* Ehrh. Affected By Harvest date

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Abstract

The variation in the content of secondary metabolites of medicinal and aromatic plants can be influenced by harvest date and climatic conditions. The objectives of this study were to evaluate the effect of seven harvest dates every two weeks (4th May, 17th May, 31st May, 14th June, 28th June, 12th July and 26th July) on the productivity of the essential oil content and composition of pennyroyal (*Mentha pulegium* L.) and apple mint (*Mentha suaveolens* Ehrh.) grown under environmental conditions of Egypt. Essential oils were extracted and the chemical composition was determined by gas chromatography - mass spectrophotometer (GC-MS). The fresh and dry biomass yields plant⁻¹ of the two mint species increased with the delay in harvesting date up to 12th and 26th July. Essential oil percentage ranged from 0.26 to 0.41% in pennyroyal and from 0.07 to 0.22 % in apple mint. The major constituents of pennyroyal oil were found to be pulegone (46.91 to 94.89%) and ρ -menthan-3-one,cis (0.38 to 48.14%). The highest relative concentration of pulegone (94.89%) was obtained at the 2nd harvest date on 17th May. The late harvest date on 26th July produced the highest relative concentration of ρ -menthan-3-one,cis (48.14 %) and the lowest concentration of pulegone (46.91%). In apple mint, the major constituents were l-linalool (58.18 to 73.27%) followed by linalyl acetate (6.10 to 24.56%), then α -terpineol (4.57 to 7.81%) and 1,8-cineole (1.72 to 4.68%). The highest relative concentrations of l-linalool (73.27%) and 1,8-cineole (4.68%) were produced from the 2nd harvest date on 17th May, while the lowest relative percentage (58.18 and 1.72 %) were recorded at the 5th harvest on 28th June. On the other hand, the highest amount of linalyl acetate (24.56%) was recorded in the 5th harvest date. It might be concluded that, the 2nd harvest date on 17th May was found to be suitable for producing the highest relative concentration of pulegone (94.89%) and l-linalool (73.27%) for pennyroyal and apple mint, respectively. While the delay in harvesting date up to 26th July increased the biosynthesis of ρ -menthan-3-one,cis (48.14 %) and linalyl acetate (24.56%) in pennyroyal and apple mint, respectively. The harvest date affects the rate of transformation of the major constituents of the essential oil of pennyroyal and apple mint plants.

Keywords: Essential oil, harvest date, *Mentha pulegium*, pulegon, *Mentha suaveolens*

INTRODUCTION

Mentha pulegium L. is one of *Mentha* species commonly known as pennyroyal. It is native to Europe, North Africa, Minor Asia and the near East [1]. Plant growth and the essential oil composition can be influenced by the ecological and climatic conditions. Aziz and Craker (2009) [2] stated that pulegone (88.05%) was the main constituent of pennyroyal oil under desert agro-system in Egypt, which is in accordance with some previously studies which revealed that pulegon was the major components in Massachusetts U.S.A (82.61%) [3], Uruguay (73.4%) [4], Bulgarian (45.4%) [5] and Portugal essential oil (35.1%) [6]. *M. pulegium* L.

poses antiseptic, antispasmodic, insect repellent, carminative, diaphoretic and anti-inflammatory activities [7], as well as antimicrobial [8], antioxidant [9], anticholinergic, anti-diabetic [10], hepatoprotective [11] and abortifacient [12] properties. Traditionally, herb was used for the treatment of fibrosis and cervical tumors [13]. The essential oil considered as possible candidate for human cancer chemotherapy [14].

Mentha suaveolens Ehrh. (apple mint or woolly mint) is native to Africa, Temperate Asia and Europe. The major constituents of apple mint differ with the chemotypes, menthyl acetate being the major components [15], dihydrocarvone [16] or

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piperitenone oxide and cis-piperitone oxide [17]. On the other hand, Aziz and Craker, 2009 [2] stated that the major constituents of apple mint were linalool (35.32%), p-menth-1-en-8-ol, (11.08%), and geranyl acetate (10.86%), respectively. *M. suaveolens* essential oil has antimicrobial [18] and antioxidant activities and can be used as a food additive to extend the shelf life of food products [19].

Harvest date affects the content and composition of secondary metabolites of medicinal and aromatic plants. The variation in the content of secondary metabolites is important for the interaction of plants with pathogens and herbivores. The concentration of terpenoids in aromatic plants varies during the day and season [20, 21]. Delaying the harvest date to the end of July promotes conversion of (–)-menthone to (–)-menthol in peppermint plant [22]. Although monoterpenes are predominant in most essential oils, many also contain sesquiterpenes. This study aimed to evaluate the effect of harvest date on the herb fresh and dry weight as well as the productivity of the essential oil content and composition of pennyroyal (*Mentha pulegium* L.), and apple mint (*Mentha suaveolens* Ehrh.) grown under environmental conditions of Egypt.

MATERIALS AND METHODS

Pennyroyal (*Mentha pulegium* L.) and apple mint (*Mentha suaveolens* Ehrh.) plants were established from cuttings originally secured from the Medicinal Plant Program in the Department of Plant Soil and Insect Sciences at the University of Massachusetts, Amherst, USA. The rhizomes (8 cm long) were rooted in sandy loam soil in a greenhouse at National Research Centre, Dokki, Cairo Egypt during February and were transplanted on 15th of March into clay pots (30 cm in diameter) filled with 12kg soil. The physical and chemical properties of the investigated soil were determined according to Cotteni *et al.*, (1982) [23].

The soil properties of the experimental pots were sandy loam (17% clay, 44.6% silt, and 38.4 % sand) with 1.18% organic matter, pH of 8.2, EC 1.0 dS/m. The available nutrition elements were 170 ppm N, 33 ppm P and 26 ppm K. At the time of transplanting the following fertilizers were added: Sulphur

fertilizer as magnesium sulphate (26.6% S) was added at the rate of 4.5 gm pot⁻¹. Nitrogen fertilizer as calcium nitrate (15.5%N) was added at the rate of 8.43 gm pot⁻¹. Phosphorus fertilizer as calcium super phosphate (16 % P₂O₅) was added at the rate of 7.2 gm pot⁻¹.

The layout of the experiment was in complete randomized design of three replicates, each replicate included 10 pots. Seven harvest dates were taken during the growing season of pennyroyal and apple mint plants on 4th May, 17th May, 31st May, 14th June, 28th June, 12th July and 26th July. The metrological data are shown in Table (1).

Fresh and dry herb weights (g plant⁻¹) were recorded and subjected to statistical analysis according to the methods of Sendecor and Cochran (1980) [24] using L.S.D at the level of 5%. Essential oil percentages of the fresh herb (100g) were determined by hydro-distillation for 3 hours using Clevenger-type apparatus, and dried over anhydrous sodium sulfate according to the Egyptian Pharmacopoeia (1984) [25]. Then, essential oil content (percentage and ml plant⁻¹) was calculated.

The essential oils were subsequently analyzed by GC-MS (Chromatography-mass spectrometry) instrument stands at the Laboratory of Medicinal and Aromatic Plants, National Research Centre with the following specifications. Instrument: aTRACE GC Ultra Gas Chromatographs (THERMO Scientific Corp., USA), coupled with a THERMO mass spectrometer detector (ISQ Single Quadrupole Mass Spectrometer). The GCMS system was equipped with a TG-WAX MS column (30 mx 0.25 mm i.d., 0.25 µm film thickness). Analyses were carried out using helium as carrier gas at a flow rate of 1.0 mL/min at a split ratio of 1:10 and the following temperature program: 60°C for 1 min; rising at 3.0 C/min to 240°C and held for 1 min. The injector and detector were held at 240°C. Diluted samples (1:10 hexane, v/v) of 0.2 µL of the mixtures were always injected. Mass spectra were obtained by electron ionization (EI) at 70 eV, using a spectral range of m/z 40-450. Most of the compounds identified using the analytical methods: mass spectra (authentic chemicals, Wiley spectral library collection and NSIT library).

Table 1. Monthly average of metrological data at the experimental area.

Month	Air temperature °C			Relative humidity %		
	Maximum	Minimum	Average	Maximum	Minimum	Average
May	45	16	25	94	6	47
June	41	18	27	88	11	51
July	41	21	29	100	9	53

RESULTS AND DISCUSSION

Data illustrated in tables (2 and 3) showed that the fresh and dry weight of herb increased with a delay in harvesting date up to 12th and 26th July) for pennyroyal and apple mint, respectively. Pennyroyal essential oil ranged from 0.26 to 0.41% and apple mint oil ranged from 0.07 to 0.22 % of all harvesting times (Tables, 2&3). The maximum essential oil percentage and content for pennyroyal plant was obtained on 28th June, while the minimum ones were recorded on 4th May date as shown in Table (2). The corresponding dates for apple mint plant were the 12th July and 4th May, respectively. Summer environmental conditions (high temperature and long days) favored essential oil biosynthesis by increasing biomass associated to plant modulation of photosynthetic carbon production into the metabolic machinery of monoterpenes biosynthesis [26].

The identified components of pennyroyal and apple mint essential oil are given in Table (4 and 5). The essential oil constituents varied with harvesting dates. Monoterpenes represented the majority (84.26 - 99.44%) of pennyroyal oil in all harvest dates i.e. α -pinene, sabinene, β - pinene, α -myrcene, dl-limonene etc.. The oxygenated compounds were dominated (82.94 - 98.36%) and consisted mainly of 3-octanol, 1,8- cineole, linalool, 3-octanol,acetate, ρ - menthan-3-one,cis, pulegone etc.. and the ρ -menthan-3-one,cis, (0.38- 48.14%). Pulegone (46.91- 94.89%) were the major component of the oxygenated compounds. In contrast, the sesquiterpenes represented a lower percentage (0.36 - 1.63%) in the oil by α -gurjunene and trans-caryophyllene whereas non-oxygenated compounds represented (1.48-2.95%).

Data in Table (4) showed that the major constituents of pennyroyal oil were found to be pulegone and ρ - menthan-3-one,cis, which ranged from 46.91 to 94.89% and from 0.38 to 48.14 %, respectively. Pulegone was found to be the major constituent and the highest relative concentration (94.89 %) was obtained at the 2nd harvest date on 17th May. The later harvest date in July 26th produced the highest relative concentration of ρ -menthan-3-one,cis (48.14 %) and the lowest concentration of pulegone (46.91%). It is clear that increasing plant age increased the biosynthesis of ρ -menthan-3-one,cis and decreased the amount of pulegone. It is well known that ρ -menthan-3-one,cis transforms to

pulegone and the environmental conditions affect the rate of transformation. These results were in accordance with some previous studies, Aziz *et al.*, 2019 [27] found that pulegone (74.43%) was the main component of *M. pulegium* oil in Egypt. Pulegone (82.61%) was the major components of pennyroyal cultivated in Massachusetts U.S.A [3], Uruguay (73.4%) [4], Bulgarian (45.4%) [5] and Portugal essential oil (35.1%) [6]. Whereas the major essential oil constituent of pennyroyal varies and can be pulegone, piperitenone-piperitone, or isomenthone-neoisomenthol, depends on the specific type of pennyroyal [28]. The major components of Bulgarian pennyroyal oil were identified as pulegone (42.9- 45.4%), piperitenone (21.7-23.1%), isomenthone (11.3-12.8%) [5]. The variations in pulegone levels could be due to the effect of ecological factors during the growing period as **described** before by Vorin *et al.*, (1990) [29] who claimed that temperature and photoperiod influenced pulegone level in mint oil.

The results shown in Table (5) clarified that the essential oil composition of *Mentha suaveolens* was characterized by a high percentage of monoterpenes (89.79 - 96.01%) with a high percentages of oxygenated compound (90.75 - 96.00 %) and the major constituents were l-linalool (58.18 to 73.27%) followed by linalyl acetate (6.10 to 24.56%), then α -terpineol (4.57 to 7.81%) and 1,8-cineole (1.72 to 4.68%). The non-oxygenated compounds ranged from 2.71 to 4.31% and the oil had lower relative concentrations of sesquiterpenes compound (2.67 - 5.69%). These results agreed with that reported by Aziz and Craker, 2009 [2] who stated that the major constituents of *Mentha suaveolens* were linalool (35.32%), ρ -menth-1-en-8-ol, (11.08%), and geranyl acetate (10.86%). The highest relative concentration of l-linalool (73.27%) and 1,8- cineole (4.68%) were produced from the 2nd harvest time on 17th May, while the lowest one (58.18 and 1.72 %) were recorded at the 5th harvest on 28th June. On the other hand, the highest relative percentage of linalyl acetate (24.56%) was recorded from the 5th harvest date. The variation in the composition of essential oil may be due to plant species and origin, and environmental conditions, especially day length and photon flux density [30].

Table 2. Fresh and dry weight and essential oil of *Mentha pulegium* L as influenced by harvest time.

Harvest times	Fresh weight g plant ⁻¹	Dry weight g plant ⁻¹	Essential oil %	Oil yield ml plant ⁻¹
4 th May	32.24	7.24	0.32	0.10
17 th May	47.60	10.65	0.35	0.17
31 st May	54.23	14.23	0.32	0.17
14 th June	54.71	13.61	0.26	0.14
28 th June	61.48	14.60	0.41	0.25
12 th July	75.41	17.61	0.32	0.24
26 th July	70.46	16.44	0.31	0.22
LSD	5.64	0.79	0.02	0.02

Table 3. Fresh and dry weight and essential oil of *Mentha suaveolens* (Ehrh.) as influenced by harvest time

Harvest times	Fresh weight g plant ⁻¹	Dry weight g plant ⁻¹	Essential oil %	Oil yield ml plant ⁻¹
4 th May	62.59	15.46	0.07	0.04
17 th May	74.05	20.51	0.09	0.07
31 st May	93.13	24.54	0.11	0.10
14 th June	101.70	26.64	0.12	0.12
28 th June	122.60	32.14	0.14	0.17
12 th July	125.39	34.30	0.22	0.28
26 th July	145.22	41.44	0.20	0.29
LSD	9.08	0.26	0.01	0.02

Table 4. The essential oil constituents of *Mentha pulegium* L as influenced by harvest time

R.T	KI	Essential oil constituents	4 th	17 th	31 st	14 th	28 th	12 th	26 th
			May	May	May	June	June	July	July
4.81	913	α-pinene	0.37	0.47	0.47	0.44	0.38	0.46	0.50
5.85	953	Sabinene	0.08	0.07	0.06	0.07	0.07	0.08	0.07
6.01	959	β-pinene	0.29	0.33	0.33	0.32	0.29	0.36	0.37
6.37	971	β-myrcen	trac.	trac.	0.02	0.01	0.06	0.03	0.03
6.83	986	3-octanol	0.43	0.13	0.11	0.12	0.16	0.07	0.03
7.64	1010	dl-limonene	0.58	0.08	trac.	0.1	0.26	0.28	0.11
7.72	1013	1,8-cineole	0.12	0.14	0.19	0.23	0.16	0.21	0.23
10.34	1083	Linalool	trac.	trac.	0.03	0.02	0.03	0.02	0.2
10.88	1095	3-octanol, acetate	trac.	0.29	trac.	0.05	0.29	0.38	0.4
12.59	1138	ρ-menthan-3-one,cis	1.08	0.38	1.45	3.22	30.52	41.97	48.14
12.96	1147	ρ-menthan-3-one,trans	trac.	0.06	0.07	0.11	1.25	1.79	1.72
13.47	1159	Isopulegone	0.95	0.72	0.93	0.84	0.67	0.66	0.57
14.27	1176	linalyl propionate	0.16	0.1	0.12	0.15	0.18	0.12	0.12
16.07	1215	Pulegone	80.20	94.89	94.76	91.64	64.28	52.97	46.91
16.88	1234	cis-isopulegone	trac.	0.11	trac.	trac.	0.11	0.01	0.02
18.09	1261	Dihydroedylan 1	trac.	0.18	0.01	trac.	0.18	0.02	0.02
22.66	1362	α-gurjunene	1.06	0.96	0.64	1.42	0.48	0.24	trac.
23.27	1375	trans-caryophllene	0.57	0.51	0.34	trac.	0.26	0.12	0.26
24.78	1408	Humulen	trac.	0.18	trac.	trac.	trac.	trac.	0.14
		Monoterpenes	84.26	97.95	98.55	97.32	98.89	99.43	99.44
		Sesquiterpenes	1.63	1.65	0.98	1.42	0.74	0.36	0.40
		Oxygenated compounds	82.94	97.00	97.67	96.38	97.83	98.22	98.36
		Non-oxygenated compounds	2.95	2.60	1.86	2.36	1.80	1.57	1.48
		Total	85.89	99.60	99.53	98.74	99.63	99.79	99.84

trac. = less than 0.005

Table 5. The essential oil constituents of *Mentha suaveolens* (Ehrh.) as influenced by harvest time

R.T	KI	Oil constituents	4 th May	17 th May	31 st May	14 th June	28 th June	12 th July	26 th July
4.8	912	α -pinene	0.08	0.13	0.13	trac.	trac.	0.1	0.5
5.85	953	Sabinene	0.07	0.13	0.12	trac.	trac.	0.11	0.12
6.01	959	β - pinene	0.18	0.3	0.29	trac.	0.05	0.25	0.27
6.3	969	β -myrcene	0.22	0.28	0.59	0.07	0.06	0.35	0.26
7.59	1009	dl- limonene	0.1	0.19	0.82	trac.	trac.	0.15	0.1
7.71	1013	1,8- cineole	3.97	4.47	4.68	2.49	1.72	4.38	3.89
8.17	1027	α -ocimene y	0.14	0.13	0.31	trac.	trac.	0.2	0.12
9.56	1064	α -terpinolene	0.12	0.16	0.26	0.06	0.06	0.19	0.13
10.2	1080	l- linalool	67.75	73.27	66.86	63.01	58.18	71.44	66.87
10.42	1085	l-octen-3-yl- acetate	trac.	1.74	2.11	1.28	1.26	1.09	0.92
10.86	1095	3-octanol,acetate	2.87	2.64	3.52	2.2	2.03	2	0.92
13.21	1153	ρ - menth-1-en-8-ol,(s)-(-)-	trac.	0.09	0.08	0.1	0.11	0.11	0.11
13.56	1161	Terpinene- 4- ol	0.06	trac.	trac.	0.05	trac.	0.05	0.05
14.21	1175	α -terpineol	6.31	trac.	7.81	4.57	4.64	5.27	4.99
15.89	1211	Geraniol	0.06	trac.	0.08	trac.	trac.	trac.	trac.
16.26	1220	Linalyl acetate	11.01	6.10	6.30	19.62	24.56	9.9	15.65
17.83	1256	Nerayl acetate	0.30	0.16	0.34	0.41	0.72	0.42	0.53
22.69	1363	α -gurjunene	0.11	0.17	0.08	0.15	0.11	0.08	0.08
23.26	1375	Trans- caryophllene	0.09	0.13	trac.	0.13	0.1	0.08	0.07
25.83	1434	Germacerene-D	1.24	2.06	0.88	2.47	2.65	1.49	1.87
26.4	1447	Bicyclogermacrene	0.06	0.11	trac.	0.11	0.1	0.07	0.07
27.54	1473	Dihidro- β -agarofuran	0.42	0.68	0.35	0.6	0.49	0.37	0.36
29.72	1524	Endo-1-bourbonanol	0.06	0.13	trac.	0.09	0.11	trac.	0.03
30.41	1541	Viridiflorol	1.23	1.77	0.91	1.25	1.41	0.77	0.76
30.61	1546	α -eudesmol	0.07	0.09	0.08	0.09	0.1	0.05	0.05
32.38	1588	Guaiol	0.4	trac.	0.46	0.46	0.52	0.28	0.25
34.22	1635	6-epishyobunol	0.07	0.13	trac.	0.11	0.1	0.05	trac.
		Monoterpenes	93.24	89.79	94.30	93.86	93.39	96.01	95.43
		Sesquiterpenes	3.75	5.27	2.76	5.46	5.69	3.24	3.54
		Oxygenated compounds	94.28	90.75	93.49	95.79	95.52	96.00	95.15
		Non-oxygenated compounds	2.71	4.31	3.57	3.53	3.56	3.25	3.82
		Total	93.88	94.9	97.06	99.32	99.08	99.25	98.97

trac. = less than 0.005

CONCLUSION

It might be concluded that the 2nd harvest time on 17th May was found to be suitable for producing the highest relative **concentration** of pulegone (94.89%) and l- linalool (73.27%) for pennyroyal and apple mint, respectively. While the delay in harvesting date up to 26th July increased the biosynthesis of ρ -menthan-3-one, cis (48.14 %) and linalyl acetate (24.56%). The harvest time affects the rate of transformation of the major constituents of the essential oil of pennyroyal and apple mint plants.

CONFLICTS OF INTEREST

There are no conflicts to declare.

REFERENCES

1. Chalchat J., Gorunovic M., Maksimovic Z. and Petrovic S., Essential oil of wild growing *Mentha*

pulegium L from Yugoslavia. *J Essential Oil Res.*, 12, 598-600 (2000).

- Aziz E.E. and Craker L.E., Essential Oil Constituents of Peppermint, Pennyroyal, and Apple Mint Grown in a Desert Agrosystem. *Journal of Herbs, Spices & Medicinal Plants*, 15(4), 361-367 (2009).
- Aziz E.E., Al-Amier H. and Craker L.E., Influence of salt stress on growth and essential oil production in peppermint, pennyroyal, and apple mint. *Journal of Herbs, Spices, and Medicinal Plants*, 14(1 & 2), 77-87 (2008).
- Lorenzo D., Paz D., Dellacassa E., Davies P., Vila R. and Canigual S., Essential oils of *Mentha pulegium* and *Mentha rotundifolia* from Uruguay. *Bras. Arch. Biol. Technol.*, 45, 519-524 (2002).
- Stoyanova A. and Georgiev E., Chemical composition of the essential oil of *Mentha pulegium* L. from Bulgaria. *J. Essent. Oil Res.*, 17, 475-476 (2005).

6. Reis-Vasco E.M.C., Coelho J.A.P. and Palavra A.M.F., Comparison of pennyroyal oils obtained by supercritical CO₂ extraction and hydrodistillation. *Flavour and Fragr. J.*,14, 156-160 (1999).
7. Marderosian A.D., Peppermint. In: Marderosian AD, ed. The review of natural products. USA: Facts and Comparisons; pp. 465-466 (2001).
8. Mahboubi M. and Haghi G., Antimicrobial activity and chemical composition of *Mentha pulegium* L. essential oil. *J. Ethnopharmacol.*,19, 325-327 (2008.).
9. El-Ghorab A.H., The chemical composition of *Mentha pulegium* L. essential oil from Egypt and its antioxidant activity. *J Essential Oil Bearing Plant*,9, 183-195 (2006.).
10. Gülçin İ., Gören A.C., Taslim P., Alwasel S.H., Kılıç O. and Bursal E., Anticholinergic, antidiabetic and antioxidant activities of Anatolian pennyroyal (*Mentha pulegium*)-analysis of its polyphenol contents by LC-MS/MS. *Biocatalysis and Agricultural Biotechnology*, 23(2020), 1-10 (2020).
11. Soares P., Assreuy A., Souza E., Lima R. and Silva T., Inhibitory effects of the essential oil of *Mentha pulegium* on the isolated rat myometrium. *Planta Med.*,71, 214-218 (2005).
12. Hameed I.H., Al-Rubaye A.F. and Kadhim M.J., Antimicrobial activity of medicinal plants and urinary tract infections. *International Journal of Pharmaceutical and Clinical Research*, 9(1), 44-50 (2017).
13. Duke J.A., *Mentha pulegium* L. (*Lamiaceae*) – Pennyroyal. In: Handbook of medicinal herbs. Boca Raton: CRC Press, Florida, USA.. pp. 307-308 (2001).
14. Shirazi F.H., Ahmadi N. and Kamalnejad M., Evaluation of Northern Iran *Mentha pulegium* L. Cytotoxicity. *Daru.*,12(3), 106-110 (2004).
15. Kokkini S. and Papageorgiou V.P, Constituents of essential oils from *Mentha x rotundifolia* growing wild in Greece. *Planta Med.* 38, 166–167 (1988).
16. Hendriks H. and Van Os F.H.L., Essential oils of two chemotypes of *M. suaveolens* during ontogenesis. *Phytochemistry*,15, 1127–1130 (1976).
17. Brada M., Bezzina M., Marlier M. and Lognay G.C., Chemical composition of the leaf oil of *Mentha rotundifolia* L. from Algeria. *J. Essent. Oil Res.*,18, 663–665 (2006).
18. Ed-Dra A., Filai F.R., Bou-Idra M., Zekkori B., Bouymajane A., Moukrad N., Benhallam F. and Bentayeb A., Application of *Mentha suaveolens* essential oil as an antimicrobial agent in fresh turkey sausages. *Journal of Applied Biology & Biotechnology*, 6(1), 7-12 (2018).
19. Bouyahya A., Belmehdi O., Abrini J., Dakka N. and Bakri Y., Chemical composition of *Mentha suaveolens* and *Pinus halepensis* essential oils and their antibacterial and antioxidant activities. *Asian Pacific Journal of Tropical Medicine*,12(3), 117-122 (2019).
20. Mrlianova M., Felklova M., Reinohl V. and Toth J., The influence of the harvest cut height on the quality of the herbal drugs *Melissae folium* and *Melissae herba*. *Planta Med.*,68, 178-180 (2002).
21. Ester R.C., Ballerini G., Sequeira A.F., Velasco G.A. and Zalazara M.F., Chemical composition of essential oil from *Tagetes minuta*. leaves and flowers. *Journal of the Argentine Chemical Society*,96(1-2), 80-86 (2008).
22. Zheljzkov V.D., Cantrell Ch.L., Astatkie T. and Ebelhar W.M., Peppermint Productivity and Oil Composition as a Function of Nitrogen, Growth Stage, and Harvest Time. *Agronomy Journal*,102(1), 124-128 (2010).
23. Cottenie A., Verloo M., Keitan L., Velghe G. and Cameriyck R., Chemical analysis of plant and soil. Lab. Of Anal. Agrochem., State Univ. Ghent-Belgium Pp. 44 – 45. (1982).
24. Sendecor G.W. and Cochran W.C., "Statistical Methods" 7th Ed., Iowa State Univ., Ames, Iowa, U.S.A., pp: 507 (1980).
25. Egyptian Pharmacopoeia, General Organization for Governmental Printing Office, *Ministry of Health, Cairo, Egypt*, pp.31-33 (1984).
26. Khanuja S.P.S., Shasany A.K., Srivastava A. and Kumar S., Assessment of genetic relationships in *Mentha* species. *Euphytica*, 111(2), 121-125 (2000).
27. Aziz E.E., Rezk A.I., Omer E.A., Nofal O.A., Salama Z.A., Fouad H. and Fouad R., Chemical composition of *Mentha pulegium* L. (Pennyroyal) plant as influenced by foliar application of different sources of zinc. *Egyptian Pharmaceutical Journal*,18(1), 53-59 (2019).
28. Croteau R. and Venkatachalam K.V., Metabolism of monoterpenes: demonstration that (+)-cis-isopulegone, not piperitenone, is the key intermediate in the conversion of (-)-isopiperitenone to (+)-pulegone in peppermint (*Mentha piperita*). *Arch Biochem. Biophys*,249, 306-315 (1986).
29. Vourin B., Brun N. and Bayte C., Effects of daylength on monoterpene composition of leaves of *Mentha x piperita*. *Phytochemistry*, 29(3), 749-755 (1990).
30. Kizil S. and Tonçer O., Influence of different harvest times on the yield and oil composition of spearmint (*Mentha spicata* L. var. *spicata*). *Journal of Food, Agriculture & Environment*, 4 (3&4), 135-137 (2006)