



Determination of Vitamins, Trace Elements, and Phytochemical Compounds in Ginkgo Biloba Leaves Extracts

Aseel Khalil Ibraheem, Mohammed Z. Thani, Mustafa Taha Mohammed

Department of Chemistry, College of Science, Mustansiriyah University, Baghdad-Iraq



CrossMark

Abstract:

The present study was conducted to extract the vitamins, trace elements and phytochemical constituents of Ginkgo biloba leaves extracts using de-ionized water and ethanol (99%) as solvents, by Soxhlet extractor. The yield of the extract by the two solvents was (12.355gm/100gm) by deionized water and was (20.525 gm/100gm) by ethyl alcohol solvent. Different types of vitamins were characterized with the above extracts using High Performance Liquid Chromatography (HPLC). Vitamins A, K, E, D3, C, B1, and B2 showed interesting results, for example, vitamin K is the most abundant in ethanol, whereas vitamin C is the most abundant in aqueous extract. These findings are motivated for deep research about the vitamins in Ginkgo biloba and their antioxidant relevance for therapeutic herbal medicine. Different metal ions, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Se, and Zn, were measured using Atomic Absorption Spectrometer (AAS). The results of the qualitative detection of the two extracts indicated the existence of Alkaloids, Steroids, Terpenes, Phenols, Carbohydrates, Glycosides, Proteins, Saponins, Tannins, and Flavonoids. Phenols and flavonoids are the major compounds exist in the plant, which are thought to be responsible about the observed antioxidant activity.

Keywords: Ginkgo Biloba; Soxhlet; Vitamins; HPLC; AAS; Phytochemical.

1. Introduction:

Medical plants are characterized by physiological active principles that have been utilized in traditional medicine years ago in the treatment of different diseases and disorders[1]. One of these medicinal plants is Ginkgo biloba, which is also one of the most studied medicinal plants[2]. Ginkgo Biloba leaves are rich in a variety of natural active ingredients and have a wide range of pharmacological activities[3], which play an important role in food[4], health care[5], medicine[6], supplements[7], anti-inflammatory, anti-cancer, antioxidant properties[8], and other fields. Ginkgo leaves are used for different purposes such as practiced antioxidant mechanisms by suppressing free radicals and reactive oxygen produced during oxidative stress[9]. The traditional methods of extracting and the determination of active substances from Ginkgo biloba include solvent extraction[10], cloud point extraction[11], accelerated solvent extraction[12], ultrasound-assisted extraction[13], supercritical fluid extraction and hydro distillation (HD)[14]. Vitamins are a disparate group of compounds; they have little in common either chemically or in their metabolic

functions. Nutritionally, they form a cohesive group of organic compounds that are required in the diet in small amounts (micrograms or milligrams per day) for the maintenance of normal health and metabolic integrity. They are thus differentiated from the essential minerals and trace elements (which are inorganic), and from essential amino and fatty acids, which are required in larger amounts[15]. Generally, the human organism cannot synthesize vitamins, and for this reason, they must be included in the human diet. The determination of vitamins plays an important role in nutritional and biochemical studies, and analytical methods are available capable of determining these vitamins in different samples that are extracted from any part of the plant [16]. Many analytical methods have been used for the analysis of vitamins: chromatographic methods[17], electromigration methods[18], microbiological assays[19], and several other methods[20]. HPLC has preferred for vitamin quantifications due to its giving more decisions and more specificity than other methods[21]. Minerals are inorganic substances that are found in all body tissues and fluids. Their presence is required for the preservation of certain

*Corresponding author ;

Receive Date: 01 June 2022, **Revise Date:** 21 June 2022, **Accept Date:** 08 August 2022, **First Publish Date:** 08 August 2022

DOI: 10.21608/EJCHEM.2022.142126.6218

©2023 National Information and Documentation Center (NIDOC)

physicochemical processes that are necessary for life, and they are chemical constituents that the body uses in a variety of ways. Even though the fact that they produce no energy, they play a crucial role in a variety of bodily functions[22]. The analysis of micronutrients in food samples is of great interest both regarding nutrition and commercial aspects. This kind of analysis is so important to know the contents of foods and environmental samples given their toxicity and essential properties[23]. The study aimed is to verify the concentrations of vitamins, metal ions, and phytochemical compounds in both two extracts, and compare the presence of these species between both, then show the role of the Ginkgo plant in the treatment of several diseases.

2. Experimental:

2.1. Collecting ginkgo Biloba leaves:

The leaves of Ginkgo biloba were collected from Sulaymaniyah north of Baghdad, Iraq, in September 2021, then washed with deionized water, dried in shade for several days at room temperature, and ground to powder.

2.2. Sample Extraction:

2.2.1. Preparation of aqueous extract:

The samples were extracted using the traditional Soxhlet procedure. A 20 g dried plant sample was refluxed in 300 mL de-ionized water for 10-12 hours. Using the Soxhlet Apparatus at a boiling water temperature. Using a rotary evaporator, the extracts were filtered and concentrated till dry. Store in a dark place for use in the next step[23].

2.2.2. Preparation of alcohol extract:

Alcohol extract was prepared in the same way as the aqueous extract separation, but using ethanol instead of water.

2.3. Determination of Vitamins concentration:

Identification of vitamins was done at the department of environment and water/Ministry of Science and Technology, using HPLC technique, (SykamS3210, Germany), with C18 column (4.6mm × 150 mm, 5µm). Mobile phase (Methanol: water) (35:65), flowrate(1mL/min), detection fluorescence system (254 nm). All the standard materials used for vitamins were pure, 99%, from Samarra/Iraq/Pharmaceuticals factory, a mixture of water: acetonitrile: glacial acetic acid was selected as a solvent for samples studied and by [94:5:1] respectively, using the mobile phase of the following mixture (Methanol: water) and by [25:75] respectively, the solution has been nominated with candidates with a diameter of 0.45µm, this combination has been chosen as a mobile phase for its suitability for the analysis of these vitamins. In the second stage, 200 mg of Ginkgo leaf extracts (aqueous and alcohol) were weighted and dissolved with a 5mL of standard solution, and then the mixture was transferred to a 25 ml volumetric flask, placed in a water bath at 0°C (65-75) °C for 10 minutes, stirring continuously until dissolved and then complete the volume with de-ionized water to the mark. Then (5 ml) from the previous extract solution was placed in a 50 mL volumetric flask, the solution was completed to the mark by deionized water, then the mixture was filtered, and the solution has been nominated with candidates with a diameter of 0.45µm before injection into the HPLC system[21][23].

2.4. Determination of trace element concentrations:

Flame Atomic Absorption Spectrophotometer (FAAS), Model AA646, Shimadzu Corporation, Kyoto/Japan, and Graphite Furnace Atomic Absorption Spectrophotometer (GFAAS) were used to determine trace element concentrations at Ghazi Hariri Hospital in Baghdad. Herbal infusions and dilutions were made with deionized water [Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Se and Zn], using a microwave digestion system before metal analysis. The microwave parameters were given in Table (1).

Table (1) Microwave procedure for digestion procedure amount of the sample [24].

Amount of sample	1 g of plant sample
Digestion reagent	digested in HNO ₃ + HClO ₄ (3:1) (20) mL
Steps	leave for one night
	heated in a water bath
	cool at room temperature
	+ 70% HClO ₄ (5 mL) heated first in a water bath for 30 minutes
	heated for 15 min until reduced the volume of the solution to 10 mL
	Diluted with deionized water to 30 mL
	filtered
	diluted with deionized water to 50 mL

A standard AAS reference stock solutions of the studied metals containing 1000 $\mu\text{g mL}^{-1}$ were diluted with (3:1) ratio of HCl: HNO_3 to prepare different concentrations of solutions (0.5, 1, 2, 5, 10 and 20) $\mu\text{g mL}^{-1}$ [24]. The standard solutions were used to generate calibration curves for the metals. Linear curves have been obtained after plotting the results. Operating conditions of spectrophotometer technique were shown in Table (2)

2.5. Phytochemical compounds:

Qualitative detection of Ginkgo extracts have been carried out to make sure there is presence of active compounds using standard procedures: alkaloids (Mayer's Test), steroids (chloroform + concentrated H_2SO_4), terpenoids (Salkowski Test), carbohydrates (Benedict's reagent), glycosides (Keller Kilianin Test), proteins (NaOH+ copper sulfate), Saponins (Foam Test), phenols, tannins (lead acetate solution), and flavonoids (Alkaline reagent Test) [25].

3. Results and Discussion:

3.1 Extraction yield:

a metric for how effective a solvent is at extracting specific components from the material, and has been defined as the number of extracts recovered by mass compared to the original amount of the sample, It is determined as follows [26]:

Mass extraction yield ($\text{g}/100\text{g} \%$) = (weight of extract/weight of Ginkgo leaves) $\times 100$

The aqueous extract yield was found to be (12.355g/100g), while alcoholic extract yield was found to be (20.525g/100g).

3.2 Vitamins:

The results were as follows Table (3) by the measurements of the HPLC system, Column C18, and wavelength (254nm) have been used with standard substances for each type of vitamin measured. The concentration per vitamin was calculated by comparing the area of the pick of the standard substance with the area of the pick for the desired vitamin and according to the following equation:

$$C (\text{sample}) = [A (\text{sample})/A (\text{standard})] \times C (\text{standard})$$

In the aqueous extract, the presence of dissolved vitamins in water (C, B2, and B1) was confirmed and the concentration of vitamin C was the highest and had the best separation of the peak at retention time (4.628 min). The result showed the presence of water-soluble vitamins at a range retention time (4-7min) at a wavelength of 254 nm Figure (1) and Table (3).

In the alcoholic extract, the presence of dissolved vitamins in fat (E, D₃, A, and K) was confirmed and the concentration of vitamin E was the highest and had the best separation of the peak at retention time (6.480 min) according to the peak of standard substances. The result showed the presence of fat-soluble vitamins at a range retention time (5-12min) at a wavelength of 254nm. Figure (2) and Table (3).

Table (2) the standard operating conditions of the Spectrophotometer

Tracelement	Cd	Co	Cr	Cu	Fe	Mn	Ni	Pb	Se	Zn
The wavelength of absorption (nm)	228.0	242.5	357.9	324.8	248.3	279.0	232.0	217.0	196.0	307.6
Lamp Current (mA)	9	10	5	5	10	4	10	9	8	7
Carrier Gas Flow (ml/min)	250	300	300	300	300	250	250	300	400	300

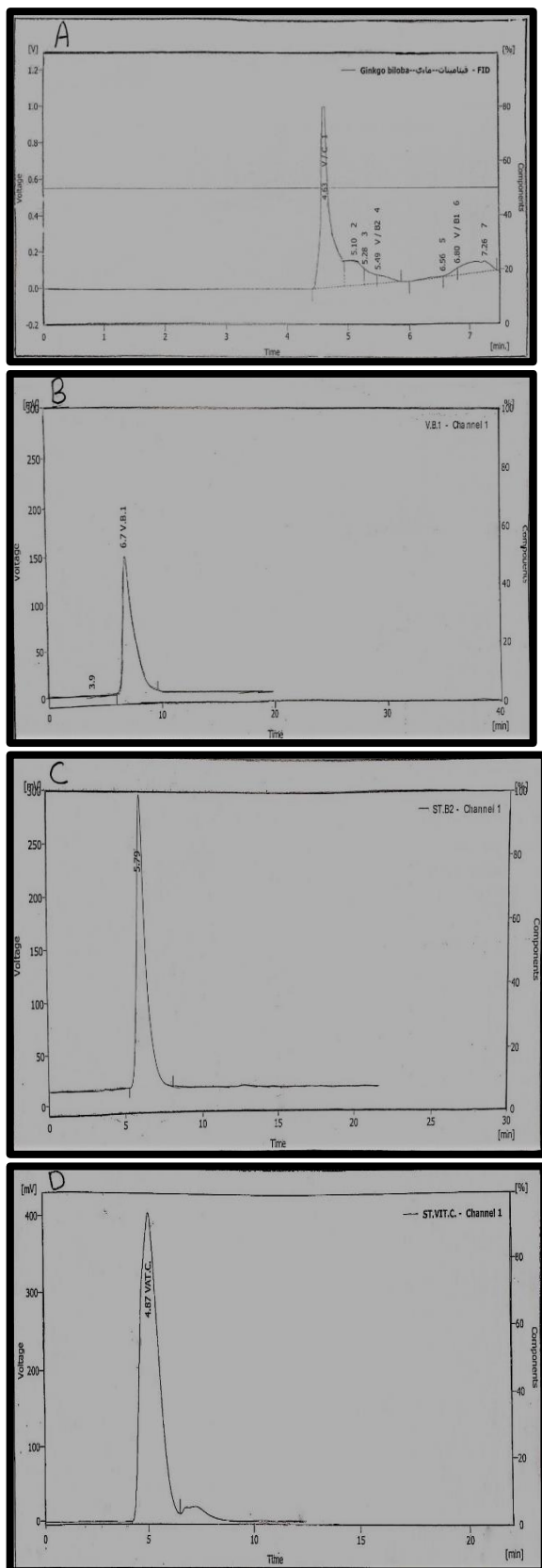


Figure (1):- HPLC analysis for (A) Vitamins water-soluble.(B) Vitamin B1 standard, (C) Vitamin B2 standard, (D) Vitamin C standard.

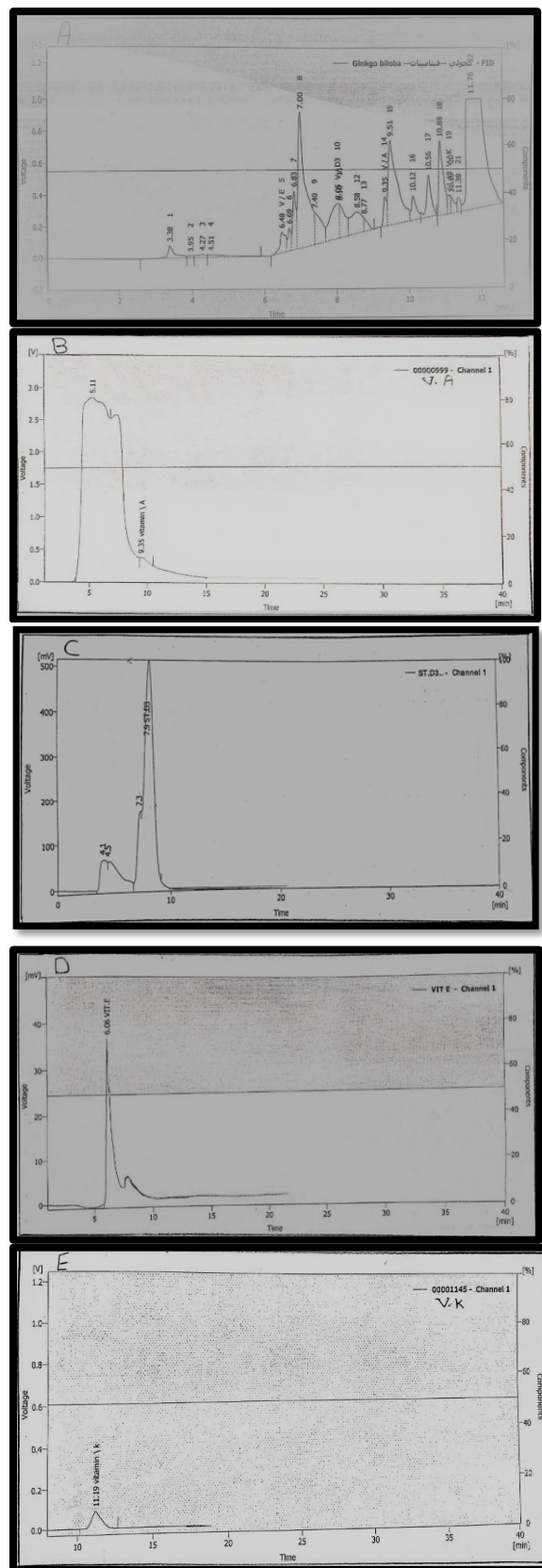


Figure (2):- HPLC analysis for (A) Vitamins fat-soluble. (B) Vitamin A standard, (C) Vitamin D3 standard, (D) Vitamin E standard, (E) Vitamin K standard

3.4. *Trace elements:* Plant samples of extracts were prepared using optimal digestion as described in the previous section? and analyzed according to the conditions are presented in Table 2).

The results of atomic absorption revealed that the presence of different amounts of trace elements necessary for human nutrition in leaves and water and alcoholic extracts Table (4).

3.5 *Phytochemical compounds:* Examining and studying the presence of plant chemical compounds in both extracts of Ginkgo Biloba leaves are shown in Table (5)

Table (3): Concentration of dissolved vitamins in the water and ethanol extracts of the Ginkgo biloba plant.

Vitamins	Resultsaqueousextract		ResultsethanolicExtract		Retention Time [Min] Extracts	Retention Time [Min] Standard
	Areaconc. [mV.s]	[$\mu\text{g.mL}^{-1}$]	Area [mV.s]	con. [$\mu\text{g.mL}$]		
A	–	–	1184.700	7.0782	9.352	9.345
E	–	–	1436.122	29.0106	6.480	6.058
D ₃	–	–	3978.072	1.8614	8.048	7.890
K	–	–	671.179	0.8663	11.132	11.188
C	10787.128	2.3412	–	–	4.628	4.873
B ₁	277.141	0.1467	–	–	6.800	6.700
.B ₂	595.259	0.2741	–	–	5.492	5.793

Table (4) Contents of trace elements in the analyzed samples of Ginkgo Biloba ($\mu\text{g/g}$)

Element	Leaves (before the extraction) ($\mu\text{g/g}$)	Aqueous extract ($\mu\text{g/g}$)	Ethyl alcohol extract ($\mu\text{g/g}$)
Cd	0.163	0.067	0.022
Co	0.264	0.095	0.087
Cr	4.221	1.552	1.399
Cu	7.936	3.499	2.130
Fe	263.72	117.881	109.263
Mn	17.553	5.456	10.535
Ni	3.836	1.572	1.233
Pb	6.092	3.216	1.671
Se	0.395	0.078	0.189
Zn	20.933	9.345	6.892

Table (5) Experiment reagents and chemical detection of Ginkgo biloba leaf extracts (water and ethanol) Active Compound

Active compounds	Experiments reagents	Indication	Water Extract	EthanolExtract
Alkaloids	Mayer's test	White precipitate	++	++
Steroids	chloroform+ concentrated H ₂ SO ₄	Layer yellow+green fluorescence	–	++
Terpenes	SalkowskiTest	Reddish-brown layer	+	++
Phenols	Lead acetate	Whiteprecipitate	++	++
Carbohydrates	Benedict's test	Green solution	++	–
Glycosides	Keller KilianinTest	brown ring	++	++
Proteins	NaOH+ copperSulphate	color	++	+
Saponins	Foam Test	Foam white	++	–
Tannins	Lead acetate	yellowish precipitate	++	++
Flavonoids	Alkaline Reagent Test	colorless	++	++

Key: ++ = High concentration

+ = Presence of bioactive compound

– = Absence of bioactive compound

All the results obtained are in agreement with previous studies.

The Ginkgo plant if it is for the acomposition of vitamins, mineral, elements, and chemical compounds [26, 32], and the varying difference in the results of this study from previous studies is due to the different habitats of plant growth used, from environmental and soil components and the quality of irrigation water, climate, temperature, and others that certainly have a significant impact on the plant.

4. Conclusions:

This study has revealed that a diet of Ginkgo Biloba can bring much health to people and further research should focus on the development of an appropriate form and route of administration of Ginkgo so that therapeutic effects are maximized.

5. Acknowledgments:

The authors would like to thank the employees at Department of Chemistry/ College of Science/ Mustansiriyah University, to facilitate the measurements and processing of the data.

6. References:

- [1] H. S. Al-Shalash, "Research Progress of Hibiscus Sabdariffa Medical Plant as Infertility Agents on Male Rabbits". *Annals of R.S.C.B.*, ISSN:1583-6258, Vol. 25, Issue 4, 2021, Pages. 11363 - 11368, Received 05 March 2021; Accepted 01 April 2021.
- [2] I. D. Boateng and X. M. Yang, "Do non-thermal pretreatments followed by intermediate-wave infrared drying affect toxicity, allergenicity, bioactive, functional groups, and flavor components of Ginkgo biloba seed? A case study," *Ind. Crops Prod.*, vol. 165, no. February, p. 113421, 2021, DOI: 10.1016/j.indcrop.2021.113421.
- [3] R. Li *et al.*, "Advances in supercritical carbon dioxide extraction of bioactive substances from different parts of Ginkgo biloba L.," *Molecules*, vol. 26, no. 13, pp. 1–16, 2021, DOI: 10.3390/molecules26134011.
- [4] C. W. Zhang, C. Z. Wang, and R. Tao, "Characterization and antioxidant activities of polysaccharides extracted from enzymatic hydrolysate of Ginkgo biloba leaves," *J. Food Biochem.*, vol. 41, no. 3, pp. 1–7, 2017, DOI: 10.1111/jfbc.12352.
- [5] H. M. R. Abdel-Latif, B. M. Hendam, M. I. Nofal, and M. A. M. El-Son, "Ginkgo biloba leaf extract improves growth, intestinal histomorphometry, immunity, antioxidant status and modulates transcription of cytokine genes in hapa-reared *Oreochromis niloticus*," *Fish Shellfish Immunol.*, vol. 117, no. June, pp. 339–349, 2021, DOI: 10.1016/j.fsi.2021.06.003.
- [6] G. Achete De Souza, S. V. De Marqui, J. N. Matias, E. L. Guiguer, and S. M. Barbalho, "Effects of Ginkgo biloba on Diseases Related to Oxidative Stress," *Planta Med.*, vol. 86, no. 6, pp. 376–386, 2020, DOI: 10.1055/a-1109-3405.
- [7] T. K. Mohanta, Y. Tamboli, and P. K. Zubaidha, "Phytochemical and medicinal importance of Ginkgo biloba L.," *Nat. Prod. Res.*, vol. 28, no. 10, pp. 746–752, 2014, DOI: 10.1080/14786419.2013.879303.
- [8] L. Zhang, T. Wu, W. Xiao, Z. Wang, G. Ding, and L. Zhao, "Enrichment and Purification of Total Ginkgo Flavonoid O-Glycosides from Ginkgo Biloba Extract with Macroporous Resin and Evaluation of Anti-Inflammation Activities In Vitro," *Molecules*, vol. 23, no. 5, 2018, DOI: 10.3390/molecules23051167.
- [9] J. Kobus-Cisowska, E. Flaczyk, M. Rudzińska, and D. Kmiecik, "Antioxidant properties of extracts from Ginkgo biloba leaves in meatballs," *Meat Sci.*, vol. 97, no. 2, pp. 174–180, 2014, DOI: 10.1016/j.meatsci.2014.01.011.
- [10] N. A. T. Alobaidy and Z. A. Thabit, "Biological Activities of *Costus speciosus* Roots," pp. 694–713, 2019.
- [11] R. Heydari and N. S. Elyasi, "Ion-pair cloud-point extraction: A new method for the determination of water-soluble vitamins in plasma and urine," *J. Sep. Sci.*, vol. 37, no. 19, pp. 2724–2731, 2014, DOI: 10.1002/jssc.201400642.
- [12] S. Saha, S. Walia, A. Kundu, K. Sharma, and R. K. Paul, "Optimal extraction and fingerprinting of carotenoids by accelerated solvent extraction and liquid chromatography with tandem mass spectrometry," *Food Chem.*, vol. 177, pp. 369–375, 2015, DOI: 10.1016/j.foodchem.2015.01.039.
- [13] H. Van Le and V. V. M. Le, "Comparison of enzyme-assisted and ultrasound-assisted extraction of vitamin C and phenolic compounds from acerola (*Malpighia emarginata* DC.) fruit," *Int. J. Food Sci. Technol.*, vol. 47, no. 6, pp. 1206–1214, 2012, DOI: 10.1111/j.1365-2621.2012.02960.x.

- [14] N. Herzi, J. Bouajila, S. Camy, M. Romdhane, and J. S. Condoret, "Comparison of different methods for extraction from *Tetraclinis articulata*: Yield, chemical composition and antioxidant activity," *Food Chem.*, vol. 141, no. 4, pp. 3537–3545, 2013, DOI: 10.1016/j.foodchem.2013.06.065.
- [15] G. Wolf, *Nutritional Biochemistry of the Vitamins*, vol. 80, no. 2. 2004.
- [16] P. Oteng, J. K. Otchere, S. Adusei, R. Q. Mensah, and E. Tei-Mensah, "Vitamin Analysis, Trace Elements Content, and Their Extractabilities in *Tetrapleura tetraptera*," *J. Chem.*, vol. 2020, 2020, DOI: 10.1155/2020/1608341.
- [17] R. Fatin Najwa and A. Azrina, "Comparison of vitamin C content in citrus fruits by titration and high performance liquid chromatography (HPLC) methods," *Int. Food Res. J.*, vol. 24, no. 2, pp. 726–733, 2017.
- [18] T. Acunha, C. Ibáñez, V. García-Cañas, C. Simó, and A. Cifuentes, "Recent advances in the application of capillary electromigration methods for food analysis and Foodomics," *Electrophoresis*, vol. 37, no. 1, pp. 111–141, 2016, doi: 10.1002/elps.201500291.
- [19] H. Fang, D. Li, J. Kang, P. Jiang, J. Sun, and D. Zhang, "Metabolic engineering of *Escherichia coli* for de novo biosynthesis of vitamin B12," *Nat. Commun.*, vol. 9, no. 1, 2018, DOI: 10.1038/s41467-018-07412-6.
- [20] Y. Zhang *et al.*, "A review of the extraction and determination methods of thirteen essential vitamins to the human body: An update from 2010," *Molecules*, vol. 23, no. 6, pp. 1–25, 2018, DOI: 10.3390/molecules23061484.
- [21] J. Mahfoud, "Using the high performance liquid chromatography (HPLC) for analysis of some vitamins," vol. 2007, pp. 21–37, 2007.
- [22] S. KO, O. C O, and O. O E, "The importance of mineral elements for humans, domestic animals and plants: A review," *African Journal of Food Science*, vol. 4, no. May, pp. 1–23, 2016.
- [23] H. İ. Ulusoy, H. Acidereli, and U. Tutar, "Optimization of Extraction Parameters for Fat Soluble Vitamins and Major Element Analysis in *Polygonum Cognatum* Meissn Plant (Madimak)," *J. Turkish Chem. Soc. Sect. A Chem.*, vol. 4, no. 1, pp. 165–165, 2016, DOI: 10.18596/jotcsa.287323.
- [24] R. E. Nwachukwu, N. I. Janefrances, I. A. Jude, I. O.-C. Fausta, U. O. Simon, and L. A. Ogechi, "Assessment of Heavy Metals Content of Some Plant-Based Medicines in Parts of Southern Nigeria, West Africa," *Int. J. Pharm. Sci. Res.*, vol. 9, no. 2, pp. 775–783, 2018, DOI: 10.13040/IJPSR.0975-8232.9(2).775-83.
- [25] Raja S and Ravindranadh K, "Preliminary phytochemical screening of different solvent extracts of the whole plant of *Acrostichum aureum*," *World J. Pharm. Res.*, vol. 2, no. 12, pp. 209–212, 2014, [Online]. Available: <http://www.wjpsonline.org/>.
- [26] E. Aspé and K. Fernández, "The effect of different extraction techniques on extraction yield, total phenolic, and anti-radical capacity of extracts from *Pinus radiata* bark," *Ind. Crops Prod.*, vol. 34, no. 1, pp. 838–844, 2011, DOI: 10.1016/j.indcrop.2011.02.002.
- [27] S. Shahidi, F. Ghahremanitamadon, S. Soleimani Asl, A. Komaki, S. Afshar, and N. Hashemi-Firouzi, "Electrophysiological, Behavioral and Molecular Study of Vitamin E and Ginkgo biloba in a Rat Model of Alzheimer's Disease," *Res. J. Pharmacogn.*, vol. 8, no. 1, pp. 39–51, 2021, DOI: 10.22127/RJP.2020.250269.1630.
- [28] B. Gafurdjanov, E. Berdiev, and U. Xoliyorov, "Study on the breeding ginkgo (ginkgo Biloba l.) in Tashkent oasis," *IOP Conf. Ser. Earth Environ. Sci.*, vol. 939, no. 1, 2021, DOI: 10.1088/1755-1315/939/1/012058.
- [29] E. D. Caldas and L. L. Machado, "Cadmium, mercury and lead in medicinal herbs in Brazil," *Food Chem. Toxicol.*, vol. 42, no. 4, pp. 599–603, 2004, DOI: 10.1016/j.fct.2003.11.004.
- [30] J. Kobus, E. Flaczyk, Z. Krejpcio, and H. Staniek, "The effect of vegetation period on contents of elements affecting the antioxidant capacity of extracts from *Ginkgo Biloba* L.," *Zywn. Nauk. Technol. Jakosc/Food. Sci. Technol. Qual.*, vol. 16, no. 4, pp. 110–115, 2009.
- [31] M. D. Luque de Castro and L. E. García-Ayuso, "Soxhlet extraction of solid materials: An outdated technique with a promising innovative future," *Anal. Chim. Acta*, vol. 369, no. 1–2, pp. 1–10, 1998, DOI: 10.1016/S0003-2670(98)00233-5.

-
- [32] Z. A. Okhti, M. E. Abdallah, and D. B. Hanna, "Phytochemical structure and Biological Effect of Ginkgo biloba leaves: A review," *Int. J. Pharm. Res.*, vol. 13, no. 02, 2021, DOI: 10.31838/ijpr/2021.13.02.180.